

TC 34

# INTERNATIONAL STANDARD 4833

INTERNATIONAL ORGANIZATION FOR STANDARDIZATION • МЕЖДУНАРОДНАЯ ОРГАНИЗАЦИЯ ПО СТАНДАРТИЗАЦИИ • ORGANISATION INTERNATIONALE DE NORMALISATION

## Microbiology — General guidance for enumeration of micro-organisms — Colony count technique at 30 °C

*Microbiologie — Directives générales pour le dénombrement des micro-organismes — Méthode par comptage des colonies obtenues à 30 °C*

First edition — 1978-02-01

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UDC 663.1

Ref. No. ISO 4833-1978 (E)

**Descriptors** : food products, animal products, microbiological analysis, counting, micro-organisms.

Price based on 4 pages

ISO 4833-1978 (E)

## FOREWORD

ISO (the International Organization for Standardization) is a worldwide federation of national standards institutes (ISO member bodies). The work of developing International Standards is carried out through ISO technical committees. Every member body interested in a subject for which a technical committee has been set up has the right to be represented on that committee. International organizations, governmental and non-governmental, in liaison with ISO, also take part in the work.

Draft International Standards adopted by the technical committees are circulated to the member bodies for approval before their acceptance as International Standards by the ISO Council.

International Standard ISO 4833 was developed by Technical Committee ISO/TC 34, *Agricultural food products*, and was circulated to the member bodies in July 1976.

It has been approved by the member bodies of the following countries :

Australia	Hungary	Portugal
Austria	India	Romania
Canada	Iran	South Africa, Rep. of
Chile	Ireland	Spain
Czechoslovakia	Israel	Turkey
France	Mexico	United Kingdom
Germany	Netherlands	U.S.A.
Ghana	Poland	Yugoslavia

The member bodies of the following countries expressed disapproval of the document on technical grounds :

New Zealand  
Thailand

# Microbiology — General guidance for enumeration of micro-organisms — Colony count technique at 30 °C

## 0 INTRODUCTION

This International Standard is intended to provide general guidance for the examination of products not dealt with by existing International Standards and for the consideration of bodies preparing reference microbiological methods of test for application to foods or to animal feeding stuffs. Because of the large variety of products within this field of application, these guidelines may not be appropriate for some products in every detail, and for some other products it may be necessary to use different methods. Nevertheless, it is hoped that in all cases every attempt will be made to apply the guidelines provided as far as possible and that deviations from them will only be made if absolutely necessary for technical reasons.

When this International Standard is next reviewed, account will be taken of all information then available regarding the extent to which the guidelines have been followed and the reasons for deviation from them in the case of particular products.

The harmonization of test methods cannot be immediate, and for certain groups of products International Standards and/or national standards may already exist that do not comply with the guidelines. In cases where International Standards already exist for the product to be tested, they should be followed, but it is hoped that when such standards are reviewed they will be changed to comply with this International Standard so that eventually the only remaining departures from these guidelines will be those necessary for well established technical reasons.

## 1 SCOPE AND FIELD OF APPLICATION

This International Standard gives general guidelines for the enumeration of micro-organisms present in products intended for human consumption or feeding of animals, by counting the colonies growing in a solid medium after incubating aerobically at 30 °C.

A limitation on the applicability of this International Standard is imposed by the method's susceptibility to a large degree of variability. The method should be applied and the results interpreted in the light of the information given in 10.2.

## 2 REFERENCES

ISO 2293, *Meat and meat products — Aerobic count at 30 °C (Reference method)*.

ISO 3565, *Meat and meat products — Detection of salmonellae (Reference method)*.

ISO . . . , *Microbiology — General guidance for preparation of dilutions*.<sup>1)</sup>

## 3 DEFINITION

For the purpose of this International Standard, the following definition applies:

**micro-organisms**: Bacteria, yeasts and moulds growing aerobically at 30 °C, under the conditions specified.

## 4 PRINCIPLE

4.1 Preparation of two poured plates, using a specified culture medium, and using a specified quantity of the test sample if the initial product is liquid, or using a specified quantity of an initial suspension in the case of other products.

Preparation of other pairs of poured plates, under the same conditions, using decimal dilutions of the test sample or of the initial suspension.

4.2 Aerobic incubation of the plates at 30 °C for 72 h.

4.3 Calculation of the number of micro-organisms per millilitre or per gram of sample from the number of colonies obtained in selected plates (see 10.1).

## 5 SAMPLING

Carry out sampling in conformity with the International Standard dealing with the product concerned.

1) In preparation.

## 6 APPARATUS AND GLASSWARE

Usual microbiological laboratory equipment, and in particular :

### 6.1 Apparatus for dry sterilization (oven) or wet sterilization (autoclave).

Apparatus that will enter into contact with the culture media, the dilution fluid or the sample, except for apparatus that is supplied sterile (particularly plastics apparatus), shall be sterilized either

- by being kept at 170 to 175 °C for not less than 1 h in an oven, or
- by being kept at 121 ± 1 °C for not less than 20 min in an autoclave.

### 6.2 Incubator, capable of being controlled at 30 ± 1 °C.

### 6.3 Petri dishes made of glass or plastics, diameter 90 to 100 mm.

### 6.4 Total delivery pipettes (blow-out pipettes), having a nominal capacity of 1 ml.

### 6.5 Water bath or similar apparatus, capable of being controlled at 45 ± 0,5 °C.

### 6.6 Colony counting equipment consisting of an illuminated base with a dark background fitted with a magnifying lens to be used at a magnification of 1,5 diameters, and a mechanical or electronic digital counter.

### 6.7 pH meter.

### 6.8 Test tubes, 18 mm × 180 mm, or flasks or bottles of suitable capacity (see 7.3 and 7.4).

## 7 CULTURE MEDIA AND DILUTION FLUID

### 7.1 Basic materials

#### **precision**

In order to improve the ~~reproducibility~~ of the results, it is recommended that, for the preparation of culture media, dehydrated basic components or complete dehydrated media should be used. The manufacturer's instructions shall be rigorously followed.

The chemicals used shall be of analytical quality.

The water used shall be distilled or deionized, and shall be free from substances that might inhibit growth of micro-organisms under the test conditions.

If the media and dilution fluid are not used immediately, they shall be kept in the dark at a temperature between

0 and + 5 °C, and in conditions that prevent any change in their composition. They shall not be kept for longer than 1 month.

### 7.2 Dilution fluid

Use a peptone-based dilution fluid containing sodium chloride, buffered or not; for example, peptone-saline dilution fluid (see 5.3 of ISO 2293) or buffered peptone water (see 6.2.1 of ISO 3565).

### 7.3 Plate count medium

#### Composition

tryptone <sup>1)</sup>	5,0 g
dehydrated yeast extract	2,5 g
anhydrous D-glucose (anhydrous dextrose)	1,0 g
agar in powder or flake form	9 to 18 g <sup>2)</sup>
water	1 000 ml

#### Preparation

Dissolve the components or the dehydrated complete medium in the water by boiling. Adjust the pH, if necessary (checking with the pH meter), so that after sterilization it is 7,0 ± 0,2 at 25 °C.

Dispense the medium into test tubes (6.8), in quantities of 15 ml per tube, or into flasks or bottles of capacity not greater than 500 ml, in quantities of approximately half the volume of the container.

Sterilize in an autoclave at 121 ± 1 °C for 20 min. If the medium is to be used immediately, cool it to 45 ± 0,5 °C in the water bath (6.5).

If not, before beginning the microbiological examination, in order to avoid any delay when pouring the medium, completely melt the medium in a boiling water bath, then cool it to 45 ± 0,5 °C in the water bath (6.5).

### 7.4 Water agar medium (if necessary — see 9.2.1.4)

#### Composition

agar in powder or flake form	9 to 18 g <sup>2)</sup>
water	1 000 ml

#### Preparation

Dissolve the agar in the water by boiling. Adjust the pH, if necessary (checking with the pH meter), so that after sterilization it is 7,0 ± 0,2 at 25 °C.

Dispense the medium into tubes (6.8), in quantities of 4 ml per tube, or into 150 ml flasks or bottles, in quantities of 100 ml per container.

Sterilize in an autoclave at 121 ± 1 °C for 20 min. If the medium is to be used immediately, cool it to 45 ± 0,5 °C in the water bath (6.5).

1) This term is only used at present by certain producers of media. Any other casein digest giving comparable results may be used.

2) According to the directions of the manufacturer.

If not, before beginning the microbiological examination, in order to avoid any delay when pouring the medium, completely melt the medium in a boiling water bath, then cool it to  $45 \pm 0,5$  °C in the water bath (6.5).

## 8 PREPARATION OF THE TEST SAMPLE

Refer to the particular International Standard dealing with the product under examination. If an International Standard is not available, it is recommended that agreement be reached on this subject by the parties concerned.

## 9 PROCEDURE

### 9.1 Test portion, initial suspension and dilutions

Refer to ISO... (see clause 2) and to the International Standard dealing with the product under examination.

Prepare the initial suspension and the dilutions using a dilution fluid meeting the requirements given in 7.2.

### 9.2 Counting technique

#### 9.2.1 Inoculation

9.2.1.1 Take two sterile Petri dishes (6.3). Using a sterile pipette (6.4), transfer to each dish 1 ml of the test sample, if liquid, or 1 ml of the initial suspension in the case of other products.

9.2.1.2 Take two other sterile Petri dishes. Using a new sterile pipette, transfer to each dish 1 ml of the 1/10 dilution (liquid product) or 1 ml of the 1/100 dilution (other products).

Repeat the procedure described in the preceding paragraph with the other dilutions.

9.2.1.3 Pour about 15 ml of the plate count medium (7.3), at  $45 \pm 0,5$  °C, into each Petri dish. The time elapsing between the end of the preparation of the initial suspension (or of the 1/10 dilution if the product is liquid) and the moment when the medium (7.3) is poured into the dishes shall not exceed 15 min.

Carefully mix the inoculum with the medium and allow the mixture to solidify, with the Petri dishes standing on a cool horizontal surface.

9.2.1.4 After complete solidification, and only in the case where it is suspected that the product under examination contains micro-organisms whose colonies will overgrow the surface of the medium, pour about 4 ml of the water agar medium (7.4), at  $45 \pm 0,5$  °C, on to the surface of the inoculated medium. Allow to solidify as described above.

This operation, if carried out, shall be mentioned in the test report.

#### 9.2.2 Incubation

Invert the prepared dishes and place them in the incubator (6.2) at  $30 \pm 1$  °C. Leave them for  $72 \pm 3$  h.

### 9.2.3 Interpretation

After the specified period of incubation (see 9.2.2), count, using the colony counting equipment (6.6), the colonies in each dish containing not more than 300 colonies.

## 10 EXPRESSION OF RESULTS

### 10.1 Method of calculation

Proceed as described in 10.1.1, 10.1.2, 10.1.3 or 10.1.4, according to the number of colonies counted in accordance with 9.2.3.

10.1.1 If one or both dishes corresponding to a certain dilution contain between 30 and 300 colonies, calculate the arithmetic mean of the number of colonies counted in the two dishes.

Retain only two significant figures, proceeding as follows :

- if the number is less than 100, round it to the nearest multiple of 5;
- if the number is greater than 100 and ends in a 5, round it to the nearest multiple of 20;
- if the number is greater than 100 and does not end in a 5, round it to the nearest multiple of 10.

Multiply this value by the reciprocal of the corresponding dilution to obtain the number of micro-organisms per millilitre or per gram of product, according to the circumstances. Express this result as a number between 1,0 and 9,9 multiplied by  $10^n$ ,  $n$  being the appropriate power of 10.

10.1.2 If there are dishes containing between 30 and 300 colonies at two consecutive dilutions, calculate the number of micro-organisms for each dilution as specified in 10.1.1, and take as the result the arithmetic mean of the two values obtained, except when the ratio of the higher value to the lower value is greater than 2; in this case, take the lower value as the result.

10.1.3 If there are fewer than 30 colonies in dishes corresponding to the test sample (liquid product) or to the initial suspension (other products), report the result as :

- fewer than 30 micro-organisms per millilitre (liquid product), or
- fewer than  $30 \times s$  micro-organisms per gram (other products), the dilution of the initial suspension being  $1/s$ .

10.1.4 If there are no colonies in dishes corresponding to the test sample (liquid product) or to the initial suspension (other products), report the result as :

- fewer than 1 micro-organism per millilitre (liquid product), or
- fewer than  $1 \times s$  micro-organisms per gram (other products), the dilution of the initial suspension being  $1/s$ .

#### 10.2 Precision of the method

For statistical reasons alone, in 95 % of cases the confidence limits of this method vary from  $\pm 12\%$  to  $\pm 37\%$  [Cowell and Morisetti (1969), *J.Sci. Fd. Agric.* **20**, 573]. In practice, even greater variation may be found especially among results obtained by different microbiologists.

#### 11 TEST REPORT

The test report shall show the method used and the results obtained. It shall also mention all operating conditions not specified in this International Standard or regarded as optional, as well as any circumstance that may have influenced the result.

The report shall include all details required for complete identification of the sample.

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