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AMENDMENT 1
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**Microbiology of the food chain —
Horizontal method for the
determination of *Vibrio* spp. —**

Part 1:
**Detection of potentially
enteropathogenic *Vibrio*
parahaemolyticus, *Vibrio cholerae*
and *Vibrio vulnificus***

AMENDMENT 1: Inclusion of
performance testing of culture media
and reagents

*Microbiologie de la chaîne alimentaire — Méthode horizontale pour
la détermination des *Vibrio* spp. —*

*Partie 1: Recherche des espèces de *Vibrio parahaemolyticus*, *Vibrio
cholerae* et *Vibrio vulnificus* potentiellement entéro-pathogènes*

*AMENDEMENT 1: Inclusion des essais de performance des milieux de
culture et réactifs*



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Microbiology of the food chain — Horizontal method for the determination of *Vibrio* spp. —

Part 1:

Detection of potentially enteropathogenic *Vibrio parahaemolyticus*, *Vibrio cholerae* and *Vibrio vulnificus*

AMENDMENT 1: Inclusion of performance testing of culture media and reagents

Clause 5

Replace the third paragraph with the following:

For performance testing of culture media and reagents, follow the procedures in accordance with ISO 11133 and Clause B.12.

5.2.1

Delete the second sentence and Table 1.

Annex B

Add the following text to the end of the annex.

B.12 Performance testing

The definition of selectivity and productivity is specified in ISO 11133. In general, follow the procedures for performance testing described in ISO 11133. Performance testing details are given in Table B.1 and Table B.2.

Table B.1 — Performance testing for the quality assurance of the culture media

Medium	Function	Incubation	Control strains	WDCM number ^a	Method of control	Criteria ^c
Alkaline saline peptone water (ASPW)	Productivity	41,5 °C ± 1 °C for 6 h ± 1 h	<i>Vibrio parahaemolyticus</i> ^d	00185	Qualitative	> 10 colonies on TCBS
			<i>Vibrio cholerae</i> ^d non-O1/non-O139 ^e	00203 ^f		
		37 °C ± 1 °C for 6 h ± 1 h	<i>Vibrio vulnificus</i> ^d	00139 00187		
Thiosulfate-citrate-bile salts (TCBS) sucrose agar	Productivity	37 °C ± 1 °C for 24 h ± 3 h	<i>Vibrio parahaemolyticus</i>	00185 ^b	Qualitative	Good growth of green colonies (2)
			<i>Vibrio furnissii</i>	00186 ^b		Good growth of yellow colonies (2)
			<i>Vibrio cholerae</i> non-O1/non-O139 ^e	00203 ^f		Good growth of yellow colonies (2)
	Selectivity	37 °C ± 1 °C for 24 h ± 3 h	<i>Escherichia coli</i> ^d	00012 00013 00090		Total inhibition (0)
Saline nutrient agar (SNA)	Productivity	37 °C ± 1 °C for 24 h ± 3 h	<i>Vibrio parahaemolyticus</i>	00185 ^b	Qualitative	Good growth (2)
			<i>Vibrio cholerae</i> non-O1/non-O139 ^e	00203 ^f		
			<i>Vibrio vulnificus</i> ^d	00139 00187		
^a World Data Centre for Microorganisms (WDCM) strain catalogue, available from: https://refs.wdcm.org/ . ^b Strain to be used as a minimum (see ISO 11133). ^c Growth is categorized as 0: no growth, 1: weak growth (partial inhibition) and 2: good growth (see ISO 11133). ^d Strain free of choice; one of the strains has to be used as a minimum (see ISO 11133). ^e The use, storage and transport of <i>V. cholerae</i> can be limited by some national restrictions and directions. ^f An appropriate sucrose-positive <i>Vibrio</i> species other than <i>V. cholerae</i> WDCM 00203 can also be used.						

Table B.2 — Performance testing of confirmation media and reagents

Medium/reagent	Function	Control strains ^a	WDCM ^b numbers	Characteristic reactions
Arginine dihydrolase saline medium	Detection of L-Arginine dihydrolase	<i>Vibrio fluvialis</i>	00137	Positive reaction: Turbidity and violet/purple colour
		<i>Vibrio parahaemolyticus</i>	00037 00185	Negative reaction: Yellow colour
Tryptone/tryptophan saline medium with indole reagent (Kovacs reagent)	Detection of indole formation from tryptophan	<i>Escherichia coli</i>	00012	Positive reaction: Formation of a red ring within 10 min
			00013	
		<i>Vibrio parahaemolyticus</i>	00090	
			00179	
			00037	
<i>Vibrio cholerae</i> non-O1/non-O139 ^d	00138			
	00185			
<i>Vibrio vulnificus</i>	00203 ^e 00139			
		<i>Enterobacter aerogenes</i>	00175	Negative reaction: Yellow/brown ring within 10 min
		<i>Citrobacter freundii</i>	00006	
		<i>Salmonella enterica</i> serovar Typhimurium ^c	00031	
Lysine decarboxylase saline medium	Detection of L-Lysine decarboxylase (LDC)	<i>Salmonella enterica</i> serovar Typhimurium ^c	00031	Positive reaction: Medium remains purple after incubation and is turbid
		<i>Salmonella enterica</i> serovar Enteritidis ^c	00030	
		<i>Enterobacter aerogenes</i>	00175	
		<i>Vibrio parahaemolyticus</i>	00185	
		<i>Proteus mirabilis</i>	00023	Negative reaction: Medium changes from purple to yellow
		<i>Citrobacter freundii</i>	00006	
		<i>Cronobacter sakazakii</i>	00214	
		<i>Cronobacter muytjensis</i>	00213	
			00012	
		<i>Escherichia coli</i>	00013	
	00090			
	00179			
<p>^a Strain free of choice; one of the strains has to be used as a minimum. The user may choose any of the strains cited for positive and negative reactions (see ISO 11133).</p> <p>^b Refer to the reference strain catalogue available on http://www.wfcc.info for information on culture collection strain numbers and contact details.^[20]</p> <p>^c Some national restrictions and directions require the use of a different serovar. Refer to national requirements relating to the choice of <i>Salmonella</i> serovars.</p> <p>^d The use, storage and transport of <i>V. cholerae</i> can be limited by some national restrictions and directions.</p> <p>^e An appropriate sucrose-positive <i>Vibrio</i> species other than <i>V. cholerae</i> WDCM 00203 can also be used.</p>				

Table B.2 (continued)

Medium/reagent	Function	Control strains ^a	WDCM ^b numbers	Characteristic reactions			
Oxidase reagent	Detection of cytochrome oxidase	<i>Pseudomonas aeruginosa</i>	00024	Positive reaction: Mauve, violet, purple or dark blue colour in the reaction time			
			00025				
			00026				
			00115				
		<i>Pseudomonas fluorescens</i>	00185	<i>Vibrio parahaemolyticus</i>	00012	Negative reaction: No colour change in the reaction time	
					00013		
					00090		
					00179		
<i>Cronobacter sakazakii</i>	00214	<i>Cronobacter mytjensis</i>	00213				
			00071	<i>Brochothrix thermosphacta</i>			
Peptone waters with different NaCl concentration: 0 %, 6 %, 10 %	Detection of halotolerance 0 % NaCl	<i>Vibrio cholerae</i> non-O1/non-O139 ^d	00203 ^e	Positive reaction: Growth (turbidity)			
			<i>Vibrio parahaemolyticus</i>	00185	Negative reaction: No growth (no turbidity)		
	Detection of halotolerance 6 % NaCl	<i>Vibrio parahaemolyticus</i>	<i>Vibrio vulnificus</i>	00037	Positive reaction: Growth (turbidity)		
				00138			
				00185			
				00139			
	Detection of halotolerance 10 % NaCl	<i>Vibrio cholerae</i> non-O1/non-O139 ^d	00203 ^e	00032	Positive reaction: Growth (turbidity)		
						00034	
	Detection of halotolerance 10 % NaCl	<i>Vibrio cholerae</i> non-O1/non-O139 ^d	00203 ^e	00037	Negative reaction: No growth (no turbidity)		
						<i>Vibrio parahaemolyticus</i>	00138
						00139	
Saline solution with toluene and β-Galactosidase reagent	Detection of β-Galactosidase	<i>Escherichia coli</i>	00012	Positive reaction: Yellow colour			
			00013				
			00090				
			00179				
		<i>Proteus mirabilis</i>	00023	<i>Vibrio parahaemolyticus</i>	00185	Negative reaction: No colour change	

^a Strain free of choice; one of the strains has to be used as a minimum. The user may choose any of the strains cited for positive and negative reactions (see ISO 11133).

^b Refer to the reference strain catalogue available on <http://www.wfcc.info> for information on culture collection strain numbers and contact details.^[20]

^c Some national restrictions and directions require the use of a different serovar. Refer to national requirements relating to the choice of *Salmonella* serovars.

^d The use, storage and transport of *V. cholerae* can be limited by some national restrictions and directions.

^e An appropriate sucrose-positive *Vibrio* species other than *V. cholerae* WDCM 00203 can also be used.