
**Textiles — Determination of antiviral
activity of textile products**

Textiles — Détermination de l'activité virucide de produits textiles

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Foreword

ISO (the International Organization for Standardization) is a worldwide federation of national standards bodies (ISO member bodies). The work of preparing International Standards is normally carried out through ISO technical committees. Each member body interested in a subject for which a technical committee has been established has the right to be represented on that committee. International organizations, governmental and non-governmental, in liaison with ISO, also take part in the work. ISO collaborates closely with the International Electrotechnical Commission (IEC) on all matters of electrotechnical standardization.

The procedures used to develop this document and those intended for its further maintenance are described in the ISO/IEC Directives, Part 1. In particular the different approval criteria needed for the different types of ISO documents should be noted. This document was drafted in accordance with the editorial rules of the ISO/IEC Directives, Part 2 (see www.iso.org/directives).

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For an explanation on the meaning of ISO specific terms and expressions related to conformity assessment, as well as information about ISO's adherence to the WTO principles in the Technical Barriers to Trade (TBT) see the following URL: Foreword - Supplementary information

The committee responsible for this document is ISO/TC 38, *Textiles*.

Introduction

Recently, along with the global improvement in the level of living, consumers are showing the trend to seek healthcare or health protective products. Also, an increase in the people's interest for protection against epidemic diseases has been noted, as the overcrowded commuting train car where the commuters experience every day, the hospitals, nursing homes, etc.

Being supported by the processing technology of textile products to provide a high performance which has been highly developed recently, the health protective and hygiene relating products have been advancing into the market.

Because those products are relatively new products and included the technical aspects out of textile technology, the testing methods have been developed by the individual producers to evaluate the product performance. That has resulted in inexistence of a unified test method, hindering for both consumers and producers a true explanation or understanding of those high functional products.

The antiviral product is one of those products and includes the technical fields of the textile technology and the biotechnology.

The demand to establish the international standard has been growing in the consumers, retailers, producers, etc. as the stakeholders in the market.

Antiviral textile products are textiles capable of reducing the number of infective virus particles that contact the surface of the textile. This standard provides a quantitative test method to assess the antiviral performance of such products.

The data obtained in objective manner by this standard give the common knowledge to all the stakeholders such as consumers, producers, retailers, etc. to understand the correct performance of the antiviral textile products.

There are two methods to quantify the number of infective virus, as infective virus titre in this standard, which are the plaque method and the TCID₅₀ method. The method used can be selected by the experience and the convenience of each testing house. Any appropriate cellular system can be used and that the testing conditions when used should be reported.

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Textiles — Determination of antiviral activity of textile products

1 Scope

This International Standard specifies testing methods for the determination of the antiviral activity of the textile products. The textile products include woven and knitted fabrics, fibres, yarns, braids, etc.

Viruses used in this International Standard are as follows:

- one of enveloped viruses, an influenza virus, which is an infective virus in humans that causes respiratory tract infection;
- one of non-enveloped viruses, a feline calicivirus, which is one of surrogates of noroviruses which are important enteric pathogens.

2 Normative references

The following documents, in whole or in part, are normatively referenced in this document and are indispensable for its application. For dated references, only the edition cited applies. For undated references, the latest edition of the referenced document (including any amendments) applies.

ISO 105-F02, *Textiles — Tests for colour fastness — Part F02: Specification for cotton and viscose adjacent fabrics*

ISO 3696, *Water for analytical laboratory use — Specification and test methods*

ISO 6330, *Textiles — Domestic washing and drying procedures for textile testing*

3 Terms and definitions

For the purposes of this document, the following terms and definitions apply.

3.1

virus

has no cell and consists of the gene material enclosed by the shell of the protein, it can replicate in the specific host cells

3.2

virus activity

ability to replicate in the specific host cells

3.3

antiviral property

property to give the morphological change or structural damage to the surface protein of virus

Note 1 to entry: As the result, the damaged virus loses the fitting to the receptor of host cell and reduces the virus activity. Depending on the type of molecules the property can also be an alteration of nucleic acids. In addition to enveloped viruses there is an alteration of envelope as well.

Note 2 to entry: It is not necessarily to imply that the change of antigenic response or the change of constituent element is the reduction of virus infectivity.

3.4

antiviral chemicals

inorganic or organic chemicals able to reduce virus activity

Note 1 to entry: The organic antiviral chemicals give the change to the surface protein of virus by the chemical adsorption. The inorganic metallic antiviral substances destroy or change the morphology of the virus by the extraction of hydrogen atom in the virus protein by OH radicals which are generated by the radical reaction.

3.5

reference cloth

cloth used to verify the stability of the test virus on a textile fabric

Note 1 to entry: The 100 % cotton cloth described in ISO 105-F02 should be used without any chemical treatments such as the fluorescent bleach, etc.

Note 2 to entry: The fabrics before the antiviral treatment may be used as a reference cloth with the same condition described in [3.5](#).

3.6

control test of specimen

test to confirm that a specimen does not affect the host cell

Note 1 to entry: This test is performed as same as actual test, but without virus.

3.7

cytopathic effect (CPE) caused by virus

effect appears as morphological change or destruction of the host cells as a result of the virus multiplication

3.8

infectivity titre of virus

number of infectious viral particles present per unit volume in a cell lysate or in a solution

3.9

plaque

lysis formed area in a cell monolayer under semisolid medium due to infection by and multiplication of a single infectious virus

3.10

plaque forming units

PFU

unit expressed as the concentration of the infectious virus per unit volume (ml)

3.11

plaque assay

assay to determine the infectivity titre of virus from PFU by using the series of dilution

3.12

TCID₅₀ method

50 % infectious dose of a wash-out virus suspension or the dilution of the virus suspension that induces a CPE in 50 % of cell culture units

Note 1 to entry: See [3.7](#).

4 Principle

The viruses are inoculated to a specimen. After specific contacting time, the remaining infectious virus is counted and the reduction rate is calculated by the comparison between the antiviral product test specimen and the reference specimen by common logarithm. There are two methods to quantify the infectious virus titre. One method is the plaque assay ([3.11](#)) and the other is the TCID₅₀ method ([3.12](#))

as explained. The selection of the method depends on the convenience and experience of the testing organization.

5 Virus and host cell

Viruses used in this standard are **an** Influenza virus and **a** feline calicivirus which is described in [Annex A](#). Moreover, the host cells are described corresponding to the viruses in [Annex A](#). One or both viruses are chosen for the test depended on the end use of the textile products.

6 Warning

This standard calls for use of the infectious viruses or substances/procedures that may be injurious to the health/environment if appropriate conditions are not observed. It refers only to technical suitability and does not absolve the user from legal obligations relating to health and safety/environment at any stage.

The warning is extended as the following. The virus in the standard shall be the one of biotechnology safety level class II classified by the directives of WHO as stated. The user of this standard shall have enough knowledge and experience of the biotechnology. Moreover, users shall comply strictly to the safety standard of the manufacturers and the domestic regulation.

7 Apparatus

- 7.1 High pressure steam sterilizer: Autoclave**, capable of operating at a temperature of $(121 \pm 2) ^\circ\text{C}$ and a pressure of $(103 \pm 5) \text{ kPa}$.
- 7.2 Dry heat sterilizer: ovens**, capable of operating at a temperature of $(180 \pm 2) ^\circ\text{C}$ and $(160 \pm 2) ^\circ\text{C}$.
- 7.3 Measuring flask**, with capacity of 1 l.
- 7.4 Scale**, with the available range of $100 \text{ g} \pm 0,1 \text{ g}$ to $0,01 \text{ g} \pm 0,0001 \text{ g}$.
- 7.5 Glass pipette**, with capacities of $50 \text{ ml} \pm 0,5 \text{ ml}$, $25 \text{ ml} \pm 0,25 \text{ ml}$, $10 \text{ ml} \pm 0,1 \text{ ml}$ and $5 \text{ ml} \pm 0,05 \text{ ml}$.
- 7.6 Plastic pipette**, with capacities of $50 \text{ ml} \pm 0,5 \text{ ml}$, $25 \text{ ml} \pm 0,25 \text{ ml}$, $10 \text{ ml} \pm 0,1 \text{ ml}$ and $5 \text{ ml} \pm 0,05 \text{ ml}$.
- 7.7 Pipetter**, capable of mounting the glass or plastic pipettes or chips.
- 7.8 Micropipette**, having the most suitable volume for each use, with a tip made of glass or plastic, and with a tolerance of 0,5 % or less.
- 7.9 Water bath**, capable of maintaining at a temperature of $(37 \pm 2) ^\circ\text{C}$, $(50 \pm 2) ^\circ\text{C}$ or $(56 \pm 2) ^\circ\text{C}$.
- 7.10 Vortex-type mixer**, used for microbial testing.
- 7.11 Freezer**, capable of operating at a temperature of $-(80 \pm 2) ^\circ\text{C}$ or $-(20 \pm 2) ^\circ\text{C}$.
- 7.12 Liquid nitrogen bath**, for the preservation approximately at $-196 ^\circ\text{C}$.
- 7.13 Membrane filter**, with a pore size of $0,22 \mu\text{m}$.
- 7.14 Refrigerator**, capable of operating at a temperature between $(2 \pm 2) ^\circ\text{C}$ and $(8 \pm 2) ^\circ\text{C}$.

7.15 **pH meter**, with a glass electrode detector.

7.16 **Inverted microscope**, capable of being used for cultured cells observation.

7.17 **Tweezers**, capable of being sterilized.

7.18 **Centrifuge**, capable of being operated at a temperature of $(4 \pm 2)^{\circ}\text{C}$, and relative centrifugal force of approximately 1 000 g.

7.19 **Biological safety cabinet**, class II.

7.20 **Vial container**, with a capacity of 30 ml and closed with the screw cap. The gasket is made of perfluoroethylene or silicone and the cap is made of polypropylene.

7.21 **96 wells microplate with the gamma radiation sterilization**, for TCID₅₀ method.



Figure 1 — 96 wells microplate for TCID₅₀ method

7.22 **6 wells plastic plate with the gamma radiation sterilization**, for plaque assay.



Figure 2 — 6 wells plastic plate for plaque assay

7.23 Flasks, for cell culture use with the gamma radiation sterilization finish, with an adherent type, a cell culture area of 75 cm² and with the vent cap, and the tight closed cap. The vent cap can be exchanged abacterial air through 0,2 µm filter.



Figure 3 — flask for cell culture use

7.24 CO₂ incubator, capable of maintaining an atmosphere with 5 % CO₂, at a temperature of (34 ± 2) °C and (37 ± 2) °C.

7.25 Incubator, capable of maintaining at a temperature of (25 ± 2) °C, (34 ± 2) °C or (37 ± 2) °C.

7.26 Centrifuge tube.

7.27 Culture container.

7.28 Test tube.

7.29 Beaker.

8 Sterilization of apparatus

Sterilize all apparatus which come in contact with the cells, the chemicals, or test specimen. The sterilization method shall be used by high pressure steam or dry heat method.

- High pressure steam sterilization: by an autoclave (7.1) at a temperature of 121 °C and a pressure of 103 kPa for 15 min.
- Dry-heat sterilization: by a dry heat sterilizer (7.2) at a temperature of 180 °C for 30 min or 160 °C for 2 h.

9 Reagent and medium

All reagents shall have the quality suitable for virological needs, ie free of toxic substances for testing microorganisms. Some of the media are available in the market.

9.1 **Water**, which must be of grade 3 according to ISO 3696.

9.2 **Eagle's minimum essential medium (EMEM)**, is available in the market. The composition is described in Annex D. If there are any components missing from the composition, add them according to the composition table.

9.3 **7,5 % sodium bicarbonate solution.**

9.3.1 Sterilize sodium bicarbonate 75 g in autoclave in a culture container with a cap closed tightly.

9.3.2 Grade 3 water is also sterilized by autoclave.

9.3.3 Dissolve sodium bicarbonate in the sterized water of 1 000 ml well.

9.4 **Formalin solution.**

9.4.1 Use for cell fixation.

9.4.2 Prepare 37 % formaldehyde solution of 100 ml,

9.4.3 Add grade 3 water of 900 ml to 9.4.2.

9.5 **Methylene blue solution**, use for the cells dyeing.

9.5.1 Prepare a measuring flask (7.3) of 1 l, then, put the following materials in the flask:

- Grade 3 water, 1 000 ml;
- Methylene blue, 0,375 g;
- 1 N sodium hydroxide solutions 62,5 µl.

9.5.2 Dissolve and mix well.

9.6 Inactivated Fetal bovine serum: FBS.

9.6.1 Put the freeze-d cryopreserved Fetal bovine serum in a package in the water bath (7.9) at a temperature of 37 °C and keep it until defrosting.

9.6.2 Then, put it in the water bath at a temperature of 56 °C and keep it for 30 min to inactivate.

9.6.3 Divide it into several tubes. Put them in the freezer (7.11) at a temperature lower than – 20 °C.

9.6.4 Just before use, put it in the water bath at a temperature of 37 °C and keep it until defrosting.

9.7 Growth medium, used for cell culture.

9.7.1 Prepare a measuring flask (7.3) of 1 l, and put the following materials into the flask:

- Grade 3 water, 800 ml;
- Kanamycin sulfate, 60 mg;
- Eagle's minimum essential medium, 9,53 g, or RPMI 1640 medium, 10,4 g.

9.7.2 Dissolve and mix well and make up whole solution to 1 000 ml by grade 3 water.

NOTE RPMI stands for Roswell Park Memorial Institute.

9.7.3 Sterilize the mixed solution of 9.7.2 by using 0,22 µm filter (7.13).

9.7.4 Add 15 ml of 7,5 % sodium bicarbonate solution (9.3) and 100 ml of the inactivated Fetal bovine serum (9.6) in the solution of 9.7.3.

NOTE When L-glutamine is not included in the EMEM purchased in the market, add it according to the composition of Annex C before use.

9.8 Maintenance medium, used for cell culture.

9.8.1 Prepare a measuring flask (7.3) of 1 l, then, put the following materials in the flask:

- Grade 3 water, 800 ml;
- Kanamycin sulfate, 60 mg;
- Eagle's minimum essential medium, 9,53 g.

9.8.2 Dissolve them well, then, make up the whole amount to 1 000 ml by adding grade 3 water.

9.8.3 Sterilize the mixed solution 9.8.2 by using the filter (7.13) with a pore size of 0,22 µm.

9.8.4 Add 15 ml of 7,5 % sodium bicarbonate solution (9.3) in the solution 9.8.3.

NOTE When L-glutamine is not included in the EMEM purchased in the market, mix it according to the composition of Annex C before use.

9.9 Double concentration of the maintenance medium 9.8.

9.9.1 Prepare a measuring flask (7.3) of 1 l, then, put the following materials in the flask:

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- Grade 3 water, 800 ml;
- Kanamycin sulfate, 120 mg;
- Eagle's minimum essential medium, 19,06 g.

9.9.2 Dissolve them well, then, make up the whole amount to 1 000 ml by adding grade 3 water.

9.9.3 Sterilize the mixed solution [9.9.2](#) by using the filter ([7.13](#)) with a pore size of 0,22 µm.

9.10 0,01 mol/l phosphate buffered saline PBS (-).

9.10.1 Prepare a measuring flask ([7.3](#)) of 1 l, then, put the following materials in the flask:

- Sodium chloride, 8 g;
- Potassium chloride, 0,2 g;
- Phosphoric acid hydrogen 2 sodium 12 hydrate, 2,9 g;
- Phosphoric acid 2 hydrogen potassium, 0,2 g.

9.10.2 Add grade 3 water by making up whole amount to 1 000 ml and dissolve well, then,

9.10.3 Sterilize the solution [9.10.2](#) by using autoclave ([7.1](#)) at a temperature of 121 °C and a pressure of 103 kPa for 15 min.

9.11 Trypsin derived from beef pancreas and PBS (-) solution.

9.11.1 Prepare a beaker, then, put the following materials in the beaker:

- 0,01 mol/l phosphate buffered saline PBS (-) ([9.10](#)), 100 ml;
- Trypsin derived from beef pancreas, 1,0 g.

9.11.2 Dissolve and mix well by using mixer for 2 h.

9.11.3 Then, sterilize the solution [9.11.2](#) by using the filter ([7.13](#)) with a pore size of 0,22 µm.

The divided solution tubes that are not used immediately are preserved in the freezer at a temperature of lower than -80 °C.

9.11.4 Prepare a test tube and put the following solutions in the test tube:

- 0,01 mol/l phosphate buffered saline PBS (-) ([9.10](#)), 9 ml;
- Trypsin derived from beef pancreas and PBS (-) mixed solution [9.11.3](#), 1ml.

9.11.5 Dissolve and mix them well.

9.11.6 Divide the solution in test tubes and preserve in the freezer at a temperature of lower than -20 °C.

9.11.7 Just before using, put it in the water bath ([7.9](#)) at a temperature of 37 °C and keep it until defrosting.

9.12 Trypsin EDTA solution.

9.12.1 Prepare a measuring flask (7.3) of 1 l, then, put the following materials in the measuring flask:

- 0,01 mol/l phosphate buffered saline PBS (-) (9.10), 1 000 ml;
- Trypsin, 2,5 g;
- Kanamycin sulfate, 0,1 g;
- Streptomycin sulfate, 0,1 g;
- Amphotericin B, 2 mg;
- EDTA, 0,014 mol.

9.12.2 Dissolve and mix well.

9.12.3 Then, sterilize the solution 9.12.2 by using the filter (7.13) with a pore size of 0,22 µm.

9.12.4 Divide the solution in test tubes and preserve in the freezer at a temperature of lower than - 20 °C.

9.12.5 Just before using, put it in the water bath (7.9) at a temperature of 37 °C and keep it until defrosting.

NOTE Trypsin EDTA solution is available in market. The products with the different components from 9.12.1, could be used after proper validation.

9.13 DEAE-dextran solution.

9.13.1 Prepare a measuring flask (7.3) of 1 l, then, put the following materials in the measuring flask:

- Grade 3 water, 1 000 ml;
- DEAE-dextran, 20 g.

9.13.2 Dissolve and mix well.

9.13.3 Sterilize the solution 9.13.2 by using the filter (7.13) with a pore size of 0,22 µm.

9.14 Agar medium, used for the plaque assay. This is prepared with A liquid and B liquid as follows and mixed well just before using.

9.14.1 A liquid.

9.14.1.1 Prepare a measuring flask (7.3) of 1 l, then, put the following materials in the measuring flask:

- Double concentration of maintenance medium (9.9), 1 000 ml;
- DEAE-dextran solution (9.13), 10 ml;
- 7,5 % sodium bicarbonate solution (9.3), 40 ml.

Mix well.

9.14.1.2 Only for the influenza virus test and for the plaque assay, add 3,0 ml of the Trypsin from beef pancreas and PBS (-) solution (9.11).

9.14.1.3 Put the solution [9.14.1.1](#) or [9.14.1.2](#) in the water bath ([7.9](#)) at a temperature of 37 °C and keep it until using.

9.14.2 B liquid.

9.14.2.1 Prepare a measuring flask ([7.3](#)) of 1 l, then, put the following materials in the measuring flask:

- Grade 3 water, 1 000 ml;
- Cell culture agar, 15 g.

Mix well.

9.14.2.2 Sterilize mixed solution [9.14.2.1](#) by using autoclave ([7.1](#)) at a temperature of 121 °C and a pressure of 103 kPa for 15 min.

9.14.2.3 Put the solution [9.14.2.2](#) in the water bath ([7.9](#)) at a temperature of 50 °C, and keep it until using.

9.15 **SCDLP medium**, used for washing-out and for suppression of agent activity of the treated textile product.

9.15.1 Prepare a measuring flask ([7.3](#)) of 1 l, then, put the following materials in the measuring flask:

- Grade 3 water, 1 000 ml;
- Peptone made of casein, 17,0 g;
- Peptone made of soybean, 3,0 g;
- Sodium chloride, 5,0 g;
- Phosphoric acid hydrogen 2 potassium, 2,5 g;
- Glucose, 2,5 g;
- Lecithin, 1,0 g.

Mix together and dissolve well, then, add,

- Nonionic surfactant, 7,0 g.

Dissolve and mix well.

9.15.2 Adjust the solution [9.15.1](#) to pH 7,0 ± 0,2 by the sodium hydroxide solution or the hydrochloric acid solution in the water bath ([7.9](#)) at a temperature of 25 °C.

9.15.3 Sterilize the mixed solution [9.15.2](#) by using autoclave ([7.1](#)) at a temperature of 121 °C and a pressure of 103 kPa for 15 min.

9.16 **Maintenance medium**, used for cell culture and for TCID₅₀ method.

9.16.1 Prepare a measuring flask ([7.3](#)) of 1 l, then, put the following materials in the measuring flask:

- Maintenance medium ([9.8](#)), 1 000 ml;
- Trypsin from beef pancreas and PBS(-) solution ([9.11](#)), 3,0 ml.

Dissolve them well.

10 Preparation

10.1 Restoration of host cell from cryopreservation

Because the host cell is cryopreserved, to defrost and then to culture are required. The procedure is as follows:

10.1.1 Put the cryopreserved host cell in the water bath (7.9) at a temperature of 37 °C and keep it for rapid defrosting.

10.1.2 Prepare a new flask for cell culture (7.23) with a vent cap lid and add 20 ml of growth medium (9.8) in the flask.

10.1.3 Put whole ampule of defrosted host cell 10.1.1 in the flask 10.1.2.

10.1.4 Put the flask 10.1.3 in the CO₂ incubator (7.24) at a temperature of 37 °C and keep it for (24 ± 2) h to culture the host cell.

10.1.5 Then, observe the flask 10.1.4 by microscope if the cells are attached on the bottom of the flask. If the growth is confirmed, then, go to next step. If not, continue to keep in the incubator.

10.1.6 Drain the remained growth medium in the flask of 10.1.5.

10.1.7 Add 20 ml of the new growth medium (9.8) to the flask of 10.1.6.

10.1.8 Put the flask 10.1.7 in the CO₂ incubator (7.24) at a temperature of 37 °C for (48 ± 2) h. Then,

10.1.9 Observe the flask of 10.1.8 by a microscope and confirm if the cells is cultured as a confluent growth on the bottom of the flask. If the growth of cells is not enough, continue the step 10.1.7 until the sufficient growth is confirmed.

10.1.10 Then, proceed the serial subcultivation by taking the following steps of 10.2.

10.2 Subculture of host cell

The host cell shall be grown by subculture. The procedure of subculture is as the following:

10.2.1 Drain an extra growth medium of the flask 10.1.9 after confirmation of a confluent growth of cells.

10.2.2 Add 5 ml of 0,01 mol/l phosphate buffered saline PBS (-) (9.11) and wash the surface of the grown cells on the bottom of the flask by the solution.

Repeat 3 times of this washing procedure.

10.2.3 Add 1 ml of Trypsin EDTA solution (9.13) in the flask of 10.2.2 and spread the solution over whole surface, then drain an extra Trypsin EDTA solution.

10.2.4 Put the flask of 10.2.3 in the CO₂ incubator (7.24) at a temperature of 37 °C for 10 min ± 1 min to keep warm. Then,

10.2.5 Observe visually the flask of 10.2.4 if the grown cells are starting to come off, if confirmed, tap the side of the flask and disperse the cells.

10.2.6 Add 5 ml of the growth medium (9.8) in the flask of 10.2.5 and pipetting the medium to make mild mix well to avoid the damage to the cells.

10.2.7 Prepare a new flask for cell culture (7.23) with a tight close cap and add 20 ml of the growth medium (9.8).

10.2.8 Add 1 ml of the cell suspension of 10.2.6 by the pipette to 10.2.7.

10.2.9 Close the cap of the flask of 10.2.8 and put the flask in the CO₂ incubator (7.24) at a temperature of 37 °C for 5 days to culture.

10.3 Cell culture for the infectious virus titre assay

The cell culture in 6 wells plate or 96 wells microplate is required for the test of the plaque assay or TCID₅₀ method.

10.3.1 Put 20 ml of growth medium (9.8) in the culture medium container (7.27) and add 1ml of the subcultured cell suspension of 10.2.6.

10.3.2 Put 3 ml of the cell suspension of 10.3.1 in each hole of the 6 wells plastic plate (7.22) for the plaque assay test.

10.3.3 Put 0,1 ml of the cell suspension of 10.3.1 in each well of the 96 wells microplate (7.21) for TCID₅₀ method.

10.3.4 Put the plate of 10.3.2 and the microplate of 10.3.3 in the CO₂ incubator (7.24) at a temperature of 37 °C and keep it for 3 days to 5 days to culture.

10.3.5 Observe the condition of the cells by the inverted microscopy if the multiplied cells are confluent.

10.4 Preparation for test virus

10.4.1 General

The viruses are cryopreserved in the freezer, so the operation to defrost and to grow them for test is required.

10.4.2 Influenza virus

10.4.2.1 Put the cryopreserved base virus in the water bath (7.9) at a temperature of 37 °C and keep it for rapid defrosting.

10.4.2.2 Drain the growth medium from the flask of 10.2.9 with the cultured cells in the monolayer.

10.4.2.3 Add 5 ml of the maintenance medium (9.8) in the flask of 10.4.2.2. Wash the surface of the cultured cells and drain the maintenance medium. Repeat the washing procedure 2 times.

10.4.2.4 Prepare a new test tube.

10.4.2.5 Put the defrosted base influenza virus in the test tube of 10.4.2.4, dilute with the maintenance medium (9.8) and adjust the concentration of virus to 10³ PFU to 10⁴ PFU or TCID₅₀ / ml.

10.4.2.6 Inoculate 1 ml of the adjusted base influenza virus of [10.4.2.5](#) on the surface of cell in the flask of [10.4.2.3](#) and spread to the whole surface.

10.4.2.7 Put the flask of [10.4.2.6](#) in the CO₂ incubator ([7.24](#)) at a temperature of 34 °C and keep it for 1 h to absorb the virus into the cells.

10.4.2.8 Put 20 ml of the maintenance medium ([9.8](#)) in the flask ([10.4.2.7](#)) and add 30 µl of Trypsin derived from beef pancreas and PBS (-) solution ([9.11](#)).

10.4.2.9 Put the flask in the CO₂ incubator ([7.24](#)) at a temperature of 34 °C for 1 to 3 days to multiply the influenza virus.

10.4.2.10 Observe the cytopathic effect by a microscope and judge the multiplication of influenza virus. If the multiplication of influenza virus is confirmed, then,

10.4.2.11 Put the multiplied virus suspension in the centrifugal tube ([7.22](#)).

10.4.2.12 Centrifuge the multiplied virus suspension of [10.4.2.11](#) by using a centrifuge at a temperature of 4 °C and 1 000 g for 15 min.

10.4.2.13 Take the supernatant suspension from the centrifugal tube after the centrifugation. This is to be the influenza virus suspension. Divide the suspension into test tubes appropriately and cryopreserve at -80 °C in the freezer ([7.11](#)).

10.4.2.14 Check the concentration of the virus if it is more than 10⁷ PFU or TCID₅₀/ml by plaque titre assay or TCID₅₀ method. If the concentration is less than 10⁷ PFU or TCID₅₀/ml, prepare it from beginning.

10.4.2.15 Just before use, put the cryopreserved virus suspension of [10.4.2.14](#) in the water bath ([7.9](#)) at a temperature of 37 °C and keep it for rapid defrosting.

10.4.2.16 This is to be the virus suspension for the test. If not use immediately, preserve in the refrigerator at a temperature of 4 °C.

10.4.3 Feline calicivirus

10.4.3.1 Put the cryopreserved base virus in the water bath ([7.9](#)) at a temperature of 37 °C and keep it for rapid defrosting.

10.4.3.2 Drain the growth medium from the flask of [10.2.9](#) with the cultured cells in the monolayer.

10.4.3.3 Add 5 ml of the maintenance medium ([9.8](#)) in the flask of [10.4.3.2](#). Wash the surface of the cultured cells and drain the maintenance medium. Repeat the washing procedure 2 times.

10.4.3.4 Prepare a new test tube.

10.4.3.5 Put the defrosted base viruses in the test tube of [10.4.3.4](#), dilute by the maintenance medium ([9.8](#)) and adjust the concentration of virus to 10⁵ PFU to 10⁶ PFU or TCID₅₀/ml.

10.4.3.6 Inoculate 1 ml of the adjusted base viruses of [10.4.3.5](#) on the surface of the cultured cells in the flask of [10.4.3.2](#) and spread to the whole surface.

10.4.3.7 Put the flask of [10.4.3.6](#) in the CO₂ incubator ([7.24](#)) at a temperature of 37 °C and keep it for 1 h to absorb the virus into the cells.

10.4.3.8 Add 20 ml of the maintenance medium ([9.8](#)) in the flask of [10.4.3.7](#).

10.4.3.9 Put the flask of [10.4.3.8](#) in the CO₂ incubator ([7.24](#)) at a temperature of 37 °C and keep it for 1 to 3 days to multiply the viruses.

10.4.3.10 Observe the cytopathic effect by a microscope and judge the multiplication of the feline calicivirus. If the multiplication of the virus is confirmed.

10.4.3.11 Put the multiplied virus suspension in the centrifugal tube.

10.4.3.12 Centrifuge the multiplied virus suspension of [10.4.3.11](#) by using the centrifuge ([7.18](#)) at a temperature of 4 °C and 1 000 g for 15 min.

10.4.3.13 Take the supernatant fluid from the centrifugal tube after the centrifugation. This is to be the virus suspension. Divide the suspension into test tubes appropriately and cryopreserve at -80 °C in the freezer ([7.11](#)).

10.4.3.14 Check the concentration of the virus if it is more than 10⁷ PFU or TCID₅₀/ml for Feline calicivirus by the plaque assay or TCID₅₀ method.

If the concentration is less than 10⁷ PFU or TCID₅₀/ml, prepare it from beginning.

10.4.3.15 Just before use, put the cryopreserved virus suspension of [10.4.3.14](#) in the water bath ([7.9](#)) at a temperature of 37 °C and keep it for rapid defrosting. This is to be a virus suspension for the test. If not use immediately, preserve in the refrigerator at a temperature of 4 °C.

10.4.4 Infectivity titre of the test viruses

The infectivity titre of the test virus shall be determined by the following procedure:

10.4.4.1 Preparation for series of the dilution for the virus suspension

10.4.4.1.1 Put 1,8 ml of the maintenance medium ([9.8](#)) in a new test tubes, keep it in the water bath with ice.

10.4.4.1.2 Add 0,2 ml of the virus suspension for the test of [10.4.2.16](#) and [10.4.3.15](#) in the test tubes of [10.4.4.1.1](#), and shake the test tubes well by Vortex mixture ([7.8](#)).

NOTE The dilution of 1/10 (10⁻¹) is prepared. The concentration of the virus suspension is 2 × 10⁶ × 10⁻¹ = 2 × 10⁵ PFU / ml or TCID₅₀/ml.

10.4.4.1.3 Put 1,8 ml of the maintenance medium ([9.8](#)) in new test tubes, keep in the water bath with ice.

10.4.4.1.4 Add 0,2 ml of the suspension of [10.4.4.1.2](#) to the test tubes of [10.4.4.1.3](#) and shake them well.

NOTE The dilution of 1/100 (10⁻²) is prepared. The concentration of the virus suspension is 2 × 10⁵ × 10⁻¹ = 2 × 10⁴ PFU / ml or TCID₅₀ / ml.

10.4.4.1.5 Repeat this procedure to prepare the series of the dilution for the virus suspension.

NOTE 1 In case of TCID₅₀ method, the series of the dilution for the virus suspension is required to prepare for the observation from the infectious wells for all of 8 wells to 0 wells.

NOTE 2 In case of the infective virus titre of 10^8 TCID₅₀/ml for the test, the virus suspension of 0,2 ml is diluted by the wash-out virus suspension of 10 ml. Then the base wash-out virus suspension becomes 2×10^6 TCID₅₀/ml. So, in case of TCID₅₀/ml, the series of the dilution for the virus suspension may be required to prepare by 10^{-7} .

10.4.4.1 Infectivity titre measurement

10.4.4.1.1 Plaque assay

Determine the infectivity titre according to [Annex B](#).

10.4.4.1.2 TCID₅₀ method

Determine TCID₅₀ according to [Annex C](#).

10.5 Preparation for test specimen

10.5.1 Control fabric

The cotton 100 % woven fabric specified by ISO 105-F02 or the untreated test sample fabric is available. Before testing, wash the reference fabric 10 times, for 10 min. at 60 °C without detergent, fluorescent bleaching agent or any other chemicals.

10.5.2 Preparation of test specimens

10.5.2.1 Obtain test specimens with mass of $0,40 \text{ g} \pm 0,05 \text{ g}$ and cut pieces with approximately 20 mm by 20 mm a piece and make up the mass with the several pieces. In case of yarns, prepare the yarns in bundle and then cut approximately 20 mm with the same mass of $0,40 \text{ g} \pm 0,05 \text{ g}$

10.5.2.2 Obtain 9 specimens for the reference cloth and 6 specimens for the antiviral test sample.

NOTE 3 reference fabric specimens and 3 antiviral test specimens are used for the control test of the effect of test specimen without virus. 3 reference specimens of the reference cloth are used for the infectivity titre measurement immediately after inoculation of virus. The remained 3 reference specimens and 3 antiviral specimens are used for the main test of this standard.

10.5.3 Sterilization of specimens

10.5.3.1 Put specimens in the vial containers one by one and put all vial containers in a metal wire basket and cover them by aluminium foil. Put caps of vial containers in the basket with wrapping of aluminium foil separately from containers.

10.5.3.2 Put the basket of [10.5.3.1](#) in the autoclave ([7.1](#)) at 121 °C and 103 kPa to sterilize for 15 min.

10.5.3.3 After sterilization, remove the foil and take out all vial containers with the specimens and put them in a safety cabinet ([7.16](#)) and keep for 60 min for cooling down, then after checking of no dew condensation in the vial containers, put the caps on all vial containers and close them.

NOTE If the high-pressure steam sterilization cannot be recommended because of the property of the antiviral agents or the characteristic of textile products, the other appropriate sterilization method could be chosen.

10.6 Control test

10.6.1 General

The purpose of the control test is to confirm the efficiency for suppression of agent activity of test specimen. The efficiency for suppression of agent activity of test specimen means no cytotoxic effect and no reduction of cell sensitivity to virus and inactivation of antiviral activity.

10.6.2 Verification of cytotoxic effect

10.6.2.1 Put 3 reference specimens and 3 antiviral test specimens sterilized in [10.5.3](#) in the vial containers, and add 20 ml of the washing out solution, SCDLP medium ([9.15](#)) in all containers. Then, put the caps on the containers and agitate them by Vortex mixer ([7.10](#)) for 5 s and 5 times.

10.6.2.2 Observe if cells damage or not, according to [Annex B](#) or [Annex C](#).

If no damage is confirmed, proceed to next step.

NOTE If the damage of cell was observed, wash-out solution should be carefully modified or changed or the amount of wash-out solution should be carefully increased.

10.6.3 Verification of cell sensitivity to virus and the inactivation of antiviral activity

10.6.3.1 Put 3 reference specimens and 3 antiviral test specimens sterilized in [Clause 10.5.3](#) in the vial containers, and add 20 ml of the washing out solution, SCDLP medium ([9.15](#)) in all containers. Then, put the caps on the containers and agitate them by Vortex mixer ([7.10](#)) for 5 s and 5 times.

10.6.3.2 Take 5 ml of washing out solution to new tubes.

10.6.3.3 Add 50 µl of virus suspension prepared to be a concentration of $(4 \text{ to } 6) \times 10^4$ PFU/ml or TCID₅₀/ml into the tubes.

10.6.3.4 Keep them at 25 °C for 30 min.

10.6.3.5 Determine infective titre according to [Annex B](#) or [Annex C](#).

10.6.3.6 A condition for verification for this test:

$\lg(\text{PFU/ml or TCID}_{50}/\text{ml of reference specimen}) - \lg(\text{PFU/ml or TCID}_{50}/\text{ml of antiviral specimen}) \leq 0,5$

NOTE If the above value is over 0,5, wash-out solution should be carefully modified or changed or the amount of wash-out solution should be carefully increased.

11 Test procedure

11.1 Preparation of specimen

All specimens are prepared in the vial containers with caps in [10.5](#).

11.2 Inoculation of virus to the specimens

Inoculate exactly 0,2 ml of the virus suspension prepared in [10.4](#) to the several points of specimen in the vial containers by pipette for all. Then put the caps on all vial containers and close them.

11.3 Contacting time

Put the vials of [11.2](#) in the incubator ([7.24](#)) and keep for 2 h as a standard time at a temperature of 25 °C.

NOTE The contacting time could be varied and might be determined by the concerned party, but not longer than 24 h.

11.4 Wash-out of virus immediately after inoculation

Immediately after inoculation of virus on 3 reference specimens in [11.2](#), add 20 ml of SCDLP ([9.15](#)) medium in the vial containers. Then, put the caps on the containers, close them and agitate them by Vortex mixer ([7.8](#)) for 5 s and 5 times for washing out the virus from the specimens.

NOTE This virus suspension is to be base wash-out virus suspension of the reference specimen.

11.5 Wash-out of virus after contacting time

After contacting for 2 h in [11.3](#), add 20 ml of SCDLP medium in the vial containers, then put the caps on the containers, close them and agitate them by Vortex mixer ([7.8](#)) for 5 s and 5 times to wash out the virus from the specimens.

NOTE These virus suspensions are to be a base wash-out virus suspension of the antiviral specimens and the reference specimens after contacting. If the virus concentration of inoculation is 10^8 (PFU/ml or TCID₅₀/ml), the concentration of the virus is to become 2×10^6 (PFU/ml or TCID₅₀/ml) (because wash-out virus suspension contains 0,2 ml virus suspension and 10 ml of the SCDLP medium, then $0,2/10 = 0,02 = 2 \times 10^{-2}$ and $10^8 \times 2 \times 10^{-2} = 2 \times 10^6$).

12 Preparation of the series of the dilution for the virus suspension

12.1 Put 1,8 ml of the maintenance medium ([9.8](#)) in new test tubes, then put and keep it in the water bath with ice.

12.2 Add 0,2 ml of the wash-out virus suspension of [11.4](#) and [11.5](#) in the test tubes of [12.1](#), and shake the test tubes well by Vortex-type mixer ([7.10](#)).

NOTE The dilution of 1/10 (10^{-1}) is prepared. The concentration of the virus suspension for this dilution is $2 \times 10^6 \times 10^{-1} = 2 \times 10^5$ PFU/ml or TCID₅₀/ml.

12.3 Put 1,8 ml of the maintenance medium ([9.8](#)) in new test tubes, then put and keep it in the water bath with ice.

12.4 Add 0,2 ml of the virus suspension of [12.2](#) to the test tubes of [12.3](#) and shake them well.

NOTE The dilution of 1/100 (10^{-2}) is prepared. The concentration of the virus suspension for this dilution is $2 \times 10^5 \times 10^{-1} = 2 \times 10^4$ PFU / ml or TCID₅₀/ml.

12.5 Repeat this procedure to prepare the series of the dilution for the virus suspension.

NOTE 1 In case of TCID₅₀ method, the series of the dilution for the virus suspension is required to prepare for observation from the infectious wells for all of 8 wells to 0 wells.

NOTE 2 In case of the infective virus titre of 10^8 TCID₅₀/ml, the virus suspension of 0,2 ml is diluted by the wash-out virus suspension of 10 ml. Then the base wash-out virus suspension becomes 2×10^6 TCID₅₀/ml. So, in case of TCID₅₀/ml, the series of the diluted virus suspension may be required by 10^{-7} .

13 Infective titre measurement

13.1 Plaque assay

Determine the infectivity titre by plaque assay according to [Annex B](#).

13.2 TCID₅₀ method

Determine TCID₅₀ according to [Annex C](#).

14 Calculation of infectivity titre

14.1 Plaque assay

$$P = Z \times R \quad (1)$$

where

P is the infective titre (PFU/0,1 ml);

Z is the arithmetic average of plaques of 2 wells (number of plaques per 0,1 ml);

R is the dilution times.

$$W = P \times 10 \quad (2)$$

where W is the infectivity titre (PFU/ml).

Then, the infectivity titre of the virus is calculated as follows:

$$V_p = W \times C$$

where

V_p is the infectivity titre (PFU/vial);

C is the wash-out virus suspension amount (ml).

14.2 TCID₅₀ method

14.2.1 The Behrens and Karber Method

In case of a logarithm dilution series.

$$Y = X \times 10^a$$

$$a = \sum p - 0,5 \tag{3}$$

where

- Y is the infective titre (TCID₅₀/0,1 ml);
- X is the dilution rate of the base virus suspension (no dilution);
- P is the ratio of the cytopathic effect at the respective dilution of the virus suspension.
- $\sum p$ is the sum of values of p .

Then, infectivity titre: A (TCID₅₀/ml) is

$$A = Y \times 10$$

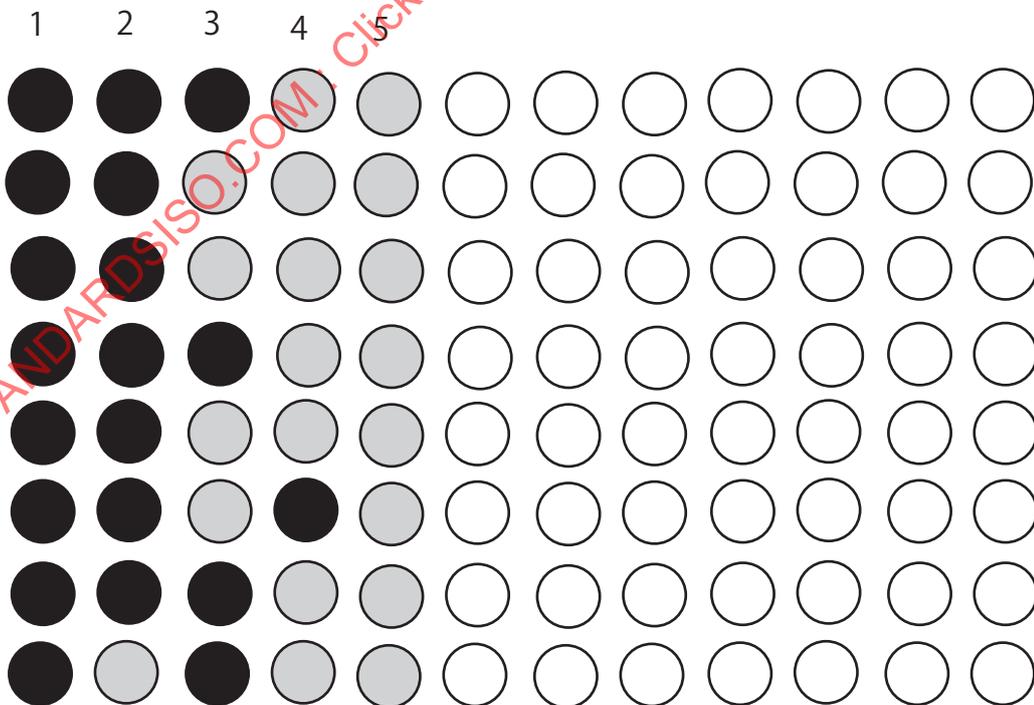
And, the infectivity titre (TCID₅₀/vial) is calculated as follows:

$$V = A \times C$$

where

- V is the infective titre (TCID₅₀/vial);
- C is the amount of wash-out virus suspension (ml);

14.2.2 Example of calculation



black: infected
 grey: not infected
 white: not inoculated

Key

- 1 inoculation column of the base virus suspension
- 2 inoculation column of 1/10 dilution of the base virus suspension
- 3 inoculation column of 1/10 dilution of the virus suspension 2
- 4 inoculation column of 1/10 dilution of the virus suspension 3
- 5 inoculation column of 1/10 dilution of the virus suspension 4

Figure 4 — Example of TCID₅₀ method

$$X = 10^0 = 1,$$

$$\sum p = 8/8 + 7/8 + 4/8 + 1/8 + 0/8 = 2,5$$

Then, $a = 2,5 - 0,5 = 2,0$

$$Y = 10^{2,0} = 1,0 \times 10^2$$

$$A = Y \times 10 = 1,0 \times 10^3$$

$$V = A \times C = A \times 20 = 1,0 \times 10^3 \times 20 = 2,0 \times 10^4$$

14.3 Test result

14.3.1 Verification of this test

- a) The virus infective titre of inoculated concentration for the test,
 - Influenza virus suspension $> 10^7$ PFU or TCID₅₀/ml
 - Feline calicivirus suspension $> 10^7$ PFU or TCID₅₀/ml
- b) To be confirmed the efficiency for suppression of agent activity of test specimen in [10.6](#).
- c) Logarithm reduction value of infective titre of reference specimen $\leq 2,0$.

$$M = \lg(Va/Vb) = \lg(Va) - \lg(Vb)$$

(4)

where

M is the reduction value;

$\lg(Va)$ is the common logarithm average of 3 infectivity titre value immediate after inoculation of the reference specimen;

$\lg(Vb)$ is the common logarithm average of 3 infectivity titre value after 2 h contacting with the reference specimen.

14.3.2 Calculation of antiviral activity value

$$M_v = \lg(V_b/V_c) = \lg(V_b) - \lg(V_c) \quad (5)$$

where

M_v is the antiviral activity value;

$\lg(V_b)$ is the common logarithm average of 3 infectivity titre value after 2 h contacting with the reference specimen;

$\lg(V_c)$ is the common logarithm average of 3 infectivity titre value after 2 h contacting with the antiviral fabric specimen.

15 Test report

The test report shall contain the following information:

- a) a reference to this International Standard;
- b) the identification of sample;
- c) the details of virus;
- d) the method to determine the infectious titre;
- e) any deviation from the specified procedures.

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Annex A (normative)

Virus strains and host cells

A.1 Virus strains and host cells

The viruses and host cells used in this standard is shown in [Table A.1](#).

Table A.1 — Viruses, host cells and media used for this standard

Virus kind	Influenza virus	Feline calicivirus
Virus strain	Influenza A virus(H3N2): A/Hong Kong/8/68: TC adapted ATCC VR-1679 Influenza A virus (H1N1): A/PR/8/34: TC adapted ATCC VR-1469,	Feline calicivirus, Strain: F-9 ATCC VR-782
Host cell ^a	MDCK cell (Dog kidney cell origin) ATCC CCL-34	CRFK cell (cat kidney cell origin) ATCC CCL-94
Growth medium ^b	EMEM (9.8.1)	RPMI 1640 (9.8.1)
^a The other host cells could be used after appropriate validation regarding to sensitivity against each viruses. ^b The other media could be used after appropriate validation for the growth of cells.		

Annex B (normative)

Infectivity titre test: Plaque assay

B.1 Test procedure

B.1.1 Pick up 6 wells plastic plate with the monolayer grown cells in the each well prepared in [Clause 10.3](#) and observe by a microscope of a confluent state of grown cells. After confirmation of the confluent state, drain extra cell growth medium from the plate.

B.1.2 Add 3 ml of the maintenance medium ([9.8](#)), wash the surface by the medium and drain the extra maintenance medium. Repeat to wash 2 times.

B.1.3 Inoculate 0,1 ml of the washing out virus suspension and the diluted virus suspension for the test in the 2 wells for one virus suspension, such as the base virus suspension into the first 2 wells and the 1/10 diluted virus suspension into the second 2 wells, as such. In the last 2 wells, inoculate the pure maintenance medium ([9.8](#)) for validation of the medium.

B.1.4 Put the plate of [B.1.3](#) in the CO₂ incubator ([7.24](#)) at the temperature listed in [Table B.1](#) and keep it for 1 h, to let the cells absorb the virus. Tilt the plate every 15 min and let to absorb the virus to whole area of the cells. Then,

B.1.5 Put 3 ml of the maintenance medium ([9.8](#)) in the plate and wash the surface, then drain the extra maintenance medium.

B.1.6 Add 3 ml of the agar medium ([9.14](#)) for the plaque assay. Close a lid and keep at room temperature for 10 min.

B.1.7 Confirm the agar coagulate, then invert the plate upside down and put it in the CO₂ incubator ([7.24](#)) at a temperature of the listed in [Table B.1](#) and keep it for 2 days to 3 days to culture. Then,

After taking it out from the CO₂ incubator ([7.24](#)), put it upright and add 3 ml of the formalin solution for cell fixation ([9.4](#)), then keep it at room temperature for more than 1 h to fix the cells. Then,

B.1.8 Drain the agar medium from [B.1.7](#), add 3 ml of the methylene blue solution ([9.6](#)), then keep it at room temperature for 15 min to dye the cells. Then,

B.1.9 Wash the extra methylene blue solution by tapped water. Confirm the dyeing of the cells. Then,

B.1.10 Count the number of plaques (white blotches),

B.1.11 Take average of two counts.

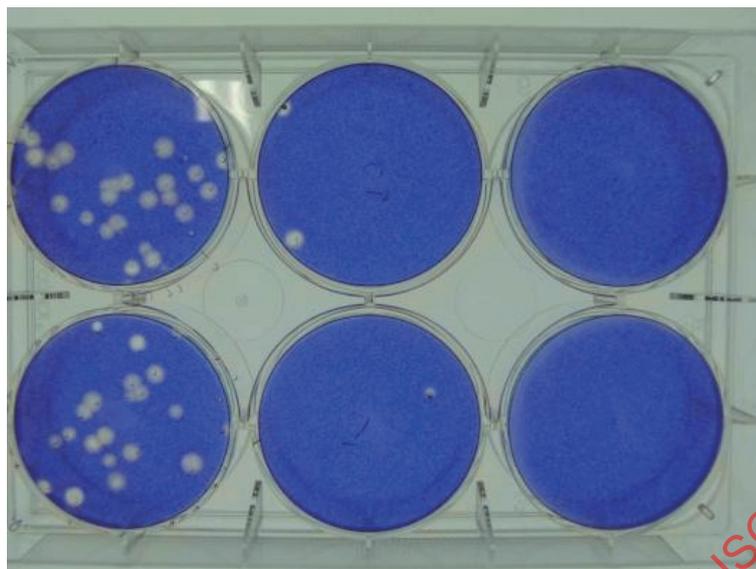


Figure B.1 — Photo of an example of Plaque assay

Table B.1 — Carbon dioxide incubator condition

Virus for examination	Influenza virus	Feline calicivirus
Clause applied	10.4.4.1.1 and 13.1	
Adsorption temperature setting °C	34	37
Culture temperature setting °C		

B.2 Determination of PFU

B.2.1 General

The plaques (white brotches) are countable by around 60 points. Over 60 points, the separation of plaques (white brotches) becomes unclear. There is another case which no plaques is observed, then as a possibility the average of two becomes less than 1. So, the determination of PFU is defined as the following in this standard.

B.2.2 Determination of PFU

As described in this standard, the plaques are counted on the dyed cells for the dilution series wells. The number of the plaques is obtained an average of two data on each dilution as [Table B.2](#).

Table B.2 — Interpretation of the data

Dilution	Wash-out virus suspension	1st dilution	2nd dilution	3rd dilution	N th dilution
Dilution rate	1	1/10	1/100	1/1 000	1/10 ^N
Average number of plaques	C1	C2	C3	C4	CN

The number of the plaques is determined in this standard as follows,

- If one of C1 to CN shows the number of plaques with 6 approximately 60, use data of 6~60 as PFU of the test,
- If C1 is 1 to less than 6, use C1 as PFU of the test,
- If C1 is less than 1, including zero, use 1 for calculation for the test.

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Annex C (normative)

Infectivity titre test: TCID₅₀ method

C.1 Test procedure

C.1.1 Pick up 96 wells microplate with the monolayer cells grown in the each well of [Clause 10.3](#) and observe by a microscope of a confluent state of the grown cells. After confirmation, drain extra growth medium from the plate.

C.1.2 Add 0,1 ml of the maintenance medium ([9.8](#)), wash the surface by the medium and drain the extra maintenance medium. Repeat to wash the surface 2 times.

C.1.3 Inoculate 0,1 ml of the wash-out virus suspension and the diluted virus suspension of the test in the 8 wells for one virus suspension, such as the base virus suspension to the first 8 wells and the 1/10 dilution for the virus suspension to the second 8 wells, as such. In the final 8 wells, inoculate the pure maintenance medium ([9.8](#)) for validation of the medium.

C.1.4 Put the microplate of [C.1.3](#) in the CO₂ incubator ([7.24](#)) at a temperature of the listed in [Table C.1](#) and keep it for 1 h, to let the cells absorb the virus. Then,

C.1.5 Drain the supernatant from the plate.

C.1.6 Add 0,1 ml of the maintenance medium ([9.8](#)), wash the surface by the medium and drain the extra maintenance medium.

C.1.7 Add 0,2 ml of the maintenance medium ([9.8](#)) for Feline calicivirus or 0,2 ml of the maintenance medium ([9.16](#)) for Influenza virus and put the microplate of [C.1.4](#) in the CO₂ incubator ([7.24](#)) at a temperature of the listed in [Table C.1](#) and keep it for 7 days to culture. Then,

C.1.8 Observe the each cell in the wells by inverted microscopy if the cytopathy of the cell is occurred. Then,

C.1.9 Confirm the cytopathy and calculate TCID₅₀ by Behrens and Karber method.

Tableau C.1 — Carbon dioxide incubator condition

Virus for test	Influenza virus	Feline calicivirus
Clause applied	10.4.4.1.2 and 13.2	
Adsorption temperature setting °C	34	37
Culture temperature setting °C		

Annex D (normative)

Composition of Media

D.1 General

The media are available in market and used for the culture of cell. As long as the culture of cell would be observed as expected, the other media could be used.

D.2 Composition of EMEM

The example of composition of EMEM is described in the [Table D.1](#). The EMEM is available in the market, however, if there are lacks of components of the composition listed in [Table D.1](#), add them accordingly to the [Table D.1](#).

Table D.1 — Composition of EMEM

Composition in 1 000 ml water		mg
Amino Acids	L-Arginine HCl	126,40
	L-Cystine 2HCl	31,20
	L-Glutamine	292,00
	L-Histidine HCl H ₂ O	41,90
	L-Isoleucine	52,50
	L-Leucine	52,50
	L-Lysine HCl	72,50
	L-Methionine	15,00
	L-Phenylalanine	32,50
	L-Threonine	47,60
	L-Tryptophan	10,00
	L-Tyrosine 2Na 2H ₂ O	51,90
	L-Valine	46,80
Vitamins	Choline Chloride	1,00
	D Calcium Pantothenate	1,00
	Folic Acid	1,00
	Myo Inositol	2,00
	Nicotinamide	1,00
	Pyridoxine HCl	1,00
	Riboflavin	0,10
Thiamine HCl	1,00	

Table D.1 (continued)

Composition in 1 000 ml water		mg
Inorganic Salts	Calcium Chloride [CaCl ₂]	200,00
	Magnesium Sulfate [MgSO ₄]	97,70
	Potassium Chloride [KCl]	400,00
	Sodium Chloride [NaCl]	6 800,00
	Sodium Phosphate Monobasic [NaH ₄ PO ₄ , H ₂ O] Monohydrate	140,00
Others	Dextrose	1 000,00
	Phenol Red Sodium Salt	10,00

D.3 Composition of RPMI 1640 medium

RPMI 1640 is also available in market. The example of composition is described in the [Table D.2](#).

Table D.2 — The composition of RPMI 1640

Composition in 1,000 ml Water		mg
Amino Acids	L-Arginine[Free Base]	200,00
	L-Asparagine[Anhydrous]	50,00
	L-Aspartic Acid	20,00
	L-Cystine 2HCl	65,20
	L-Glutamic Acid	20,00
	L-Glutamine	300,00
	Glycine	10,00
	L-Histidine[Free Base]	15,00
	Hydroxy-L-Proline	20,00
	L-Isoleucine	50,00
	L-Leucine	50,00
	L-Lysine HCl	40,00
	L-Methionine	15,00
	L-Phenylalanine	15,00
	L-Proline	20,00
	L-Serine	30,00
	L-Threonine	20,00
	L-Tryptophan	5,00
L-Tyrosine 2Na 2H ₂ O	28,83	
L-Valine	20,00	

Table D.2 (continued)

Composition in 1,000 ml Water		mg
Vitamines	Biotin	0,20
	Choline Chloride	3,00
	Folic Acid	1,00
	myo-Inositol	35,00
	Niacinamide	1,00
	D-Pantothenic Acid Hemicalcium	0,25
	p-Aminobenzoic Acid	1,00
	Pyridoxine HCl	1,00
	Riboflavin	0,20
	Thiamine HCl	1,00
	Vitamine B12	0,005
Inorganic Salts	Calcium Nitrate 4H ₂ O	100,00
	Magnesium Sulfate[Anhydrous]	48,84
	Potassium Chloride	400,00
	Sodium Chloride	6 000,00
	Sodium Phosphate Dibasic[Anhydrous]	800,00
Others	D-Glucose	2 000,00
	Glutathione, Reduced	1,00
	Phenol Red Na	5,30

Annex E (informative)

Additional virus: Polio virus

E.1 General

Poliovirus is selected as test virus because it has a high resistance to chemicals, is acid-stable and is unaffected by lipid solvents such as ether, and most detergents or quarternary products.

Polio virus is tinier virus than influenza or calicivirus. Main flame of this standard chooses two viruses as influenza in category of with envelope and feline calicivirus in category of without envelope, but polio virus in category of without envelope has been also developed in parallel. As information, testing method for polio virus is described in this annex.

E.2 Polio virus strain and host cell

Table E.1 — Polio virus strain and host cell

Virus kind	Polio virus
Virus strain	Poliovirus type1 LSc-2ab (This is a live oral poliovirus vaccine strain which complies with the WHO standard)
Host cell	HEp-2 cell (human pharynx cancer cell origin) ATCC CCL-23
Culture medium	EMEM (9.8.1)

E.3 Condition of test

All conditions for this test procedure are exactly same as for feline calicivirus.

Annex F (informative)

Testing method using SPF embryonated hen's eggs

F.1 General

The embryonated hen's has been used in the traditional biotechnology testing method, especially for the production of the vaccine for epidemic disease caused by infectious viruses. As stated in this standard, the technology to use a cultured cell has been becoming more common method to biotechnology testing to avoid the use of living body such as an embryonated hen. However, this method is still existing and used in some testing houses, so this technology is described in this annex.

F.2 Scope

This method specifies to testing method for antiviral activity assay of the textile products by using SPF embryonated hen's eggs.

F.3 Terms and definitions

F.3.1 SPF embryonated hen's eggs

eggs without experience to be infected by the pathogenic microorganism used for production of vaccine and used through this standard. SPF stands for specific pathogen free.

F.4 Preparation of specimen

F.4.1 Take a specimen with the weight of 0,2 g each and take 3 specimens from one sample.

F.4.2 For the sheet like sample, such as woven, knitted fabric or nonwoven fabric, cut 0,2 g specimen into 1,5 cm square and pile up them.

F.4.3 Put the specimen into the polyethylene bag with vinyl zipper.

NOTE No need to sterilization of specimen.

F.5 Preparation of the embryonated hen's eggs

Choose 10 day old embryonated hen's eggs.

F.6 Preparation of virus

Influenza virus: chick-embryo-adapted strain of A/PR/8/34 (H1N1) or A/Aichi/3/68 (H3N2) is used in this test.

F.6.1 Virus cultivation

F.6.1.1 Inoculate a 0,2 ml-ml aliquot of the virus suspension into allantoic cavity of the eggs.

F.6.1.2 Incubate the eggs in the incubator at a temperature of 35 °C, for 3 days.

F.6.1.3 After incubation, put the eggs into refrigerator to cool down overnight, and then take the allantoic fluid from the eggs.

F.6.1.4 Examine the all fluids taken from the eggs to confirm multiplication of influenza virus. If infected, the haemagglutination is observed. If confirmed, the fluids are considered as virus fluids, or virus media.

F.6.1.5 Just before testing, dilute the virus fluids with the phosphate buffered saline solution (PBS) (pH 7.2) to make the concentration of virus as 10^7 EID₅₀/0,2 ml.

F.6.2 Preparation of 0,5 % chicken red blood cell suspension

F.6.2.1 Draw a 4,0 ml aliquot of chicken venous blood by using a syringe which is containing 1,0 ml of the sterilized 2 % sodium citrate, and put it into a test tube.

F.6.2.2 Mount the test tube on the centrifuge and centrifugalize it by 1 000 g for 10 min.

F.6.2.3 Drain the supernatant and then add a 5 ml aliquot of PBS to the test tube.

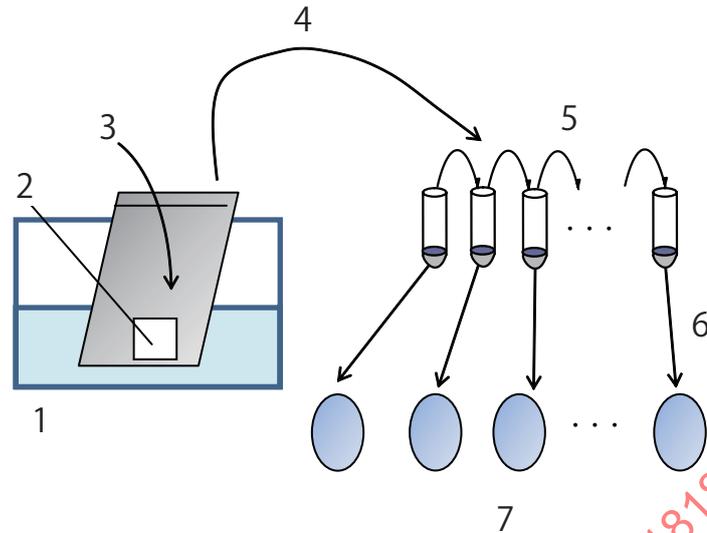
F.6.2.4 Repeat to centrifuge another two times. Then,

F.6.2.5 Mix 0,5 ml aliquot of the settled out chicken red blood and 99,5 ml aliquot of PBS and shake the test tube well.

F.7 Test procedure

F.7.1 General

General process image is shown in [Figure F.1](#).

**Key**

- 1 water bath 4 °C
- 2 specimen in a plastic bag, 3
- 3 plastic bag with virus suspension
- 4 recover virus suspension after contacting
- 5 dilution series of virus suspension by PBS
- 6 inoculation of virus suspension in the eggs
- 7 eggs for incubation

Figure F.1 — General process image of embryonated eggs method

NOTE Regarding to [Figure F.1.1](#), open air in room with 25 °C could be used alternatively.

F.7.2 Contact the virus suspension to the specimen

F.7.2.1 Put the specimen in a polyethylene bag with zipper.

F.7.2.2 Put the virus solution diluted by PBS into the polyethylene bag with the specimen.

F.7.2.3 Put the bags into the water bath with 4 °C or place the bags in open air in room with 25 °C and keep for 10 min or 2 hr as contacting time.

F.7.3 Inoculation of the reacted virus suspension to 10 day old embryonated hen's eggs

F.7.3.1 Recover the reacted virus suspension from the plastic bag of [F.7.2.3](#).

F.7.3.2 Dilute the reacted virus suspension by series of dilution, such as 10^{-1} , 10^{-2} , 10^{-3} , etc.

F.7.3.3 Inoculate the diluted virus suspension of 0,2 ml aliquot into allantoic cavity of the eggs, three eggs for each dilution.

F.7.3.4 Incubate them for two days at 35 °C.

F.7.4 Virus assay

F.7.4.1 After two days incubation, take out the allantoic fluid from the eggs and put them into test tubes. Then,

F.7.4.2 Add 0,5 % chicken red blood cell suspension into the test tubes.

F.7.4.3 Observe the test tubes if the haemagglutination is found or not.

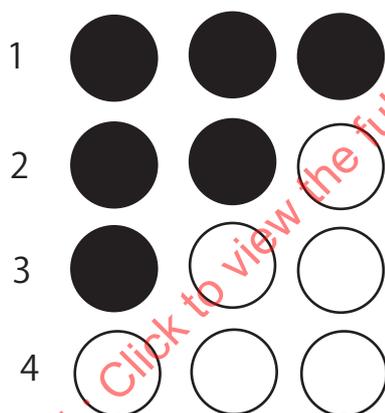
- Virus infection (-): Red cell precipitated is observed but no condensation.
- Virus infection (+): Red cell condensation observed.

F.7.5 Calculation of the virus titre

Calculate the virus titre EID_{50} by the method of Reed and Muench.

50 % EID is calculated from the cumulative infected number of each dilution at 50 %.

The example of calculation is shown in the [Figure F.2](#).



Key

- 1 inoculation row of the virus suspension dilution 10^{-4}
- 2 inoculation row of the virus suspension dilution 10^{-5}
- 3 inoculation row of the virus suspension dilution 10^{-6}
- 4 inoculation row of the virus suspension dilution 10^{-7}

- Black infected
- White not infected

Figure F.2 — Example of the embryonated eggs method

In the example,