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**Biological evaluation of medical devices —**  
**Part 5:**  
Tests for *in vitro* cytotoxicity

*Évaluation biologique des dispositifs médicaux —*  
*Partie 5: Essais concernant la cytotoxicité in vitro*



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## Foreword

ISO (the International Organization for Standardization) is a worldwide federation of national standards bodies (ISO member bodies). The work of preparing International Standards is normally carried out through ISO technical committees. Each member body interested in a subject for which a technical committee has been established has the right to be represented on that committee. International organizations, governmental and non-governmental, in liaison with ISO, also take part in the work. ISO collaborates closely with the International Electrotechnical Commission (IEC) on all matters of electrotechnical standardization.

International Standards are drafted in accordance with the rules given in the ISO/IEC Directives, Part 3.

Draft International Standards adopted by the technical committees are circulated to the member bodies for voting. Publication as an International Standard requires approval by at least 75 % of the member bodies casting a vote.

International Standard ISO 10993-5 was prepared by Technical Committee ISO/TC 194, *Biological evaluation of medical devices*.

This second edition cancels and replaces the first edition (ISO 10993-5:1992), which has been technically revised.

ISO 10993 consists of the following parts, under the general title *Biological evaluation of medical devices*:

- *Part 1: Evaluation and testing*
- *Part 2: Animal welfare requirements*
- *Part 3: Tests for genotoxicity, carcinogenicity and reproductive toxicity*
- *Part 4: Selection of tests for interactions with blood*
- *Part 5: Tests for in vitro cytotoxicity*
- *Part 6: Tests for local effects after implantation*
- *Part 7: Ethylene oxide sterilization residuals*
- *Part 8: Guidance for reference materials*
- *Part 9: Framework for the identification and quantification of potential degradation products*
- *Part 10: Tests for irritation and sensitization*
- *Part 11: Tests for systemic toxicity*
- *Part 12: Sample preparation and reference materials*
- *Part 13: Identification and quantification of degradation products from polymers*
- *Part 14: Identification and quantification of degradation products from ceramics*
- *Part 15: Identification and quantification of degradation products from metals and alloys*
- *Part 16: Toxicokinetic study design for degradation products and leachables*
- *Part 17: Methods for establishment of allowable limits for leachable substances using health-based risk assessment*
- *Part 18: Chemical characterization*

Future parts will deal with other relevant aspects of biological testing.

## Introduction

Due to the general applicability of *in vitro* cytotoxicity tests and their widespread use in evaluating a large range of medical devices and materials, it is the purpose of this part of ISO 10993, rather than to specify a single test, to define a scheme for testing which requires decisions to be made in a series of steps. This should lead to the selection of the most appropriate test.

Three categories of test are listed: extract test, direct-contact test, indirect-contact test.

The choice of one or more of these categories depends upon the nature of the sample to be evaluated, the potential site of use and the nature of the use.

This choice then determines the details of the preparation of the samples to be tested, the preparation of the cultured cells, and the way in which the cells are exposed to the samples or their extracts.

At the end of the exposure time, the evaluation of the presence and extent of the cytotoxic effect is undertaken. It is the intention of this part of ISO 10993 to leave open the choice of type of evaluation. Such a strategy makes available a battery of tests, which reflects the approach of many groups which advocate *in vitro* biological tests.

The numerous methods used and end-points measured in cytotoxicity determination can be grouped into categories of evaluation type:

- a) assessments of cell damage by morphological means;
- b) measurements of cell damage;
- c) measurements of cell growth;
- d) measurements of specific aspects of cellular metabolism.

There are, therefore, several alternative means of producing results in each of these four categories. The investigator should be aware of the categories of test and into which a particular technique fits, in order that comparisons may be made with other results on similar medical devices or materials, and in order that interlaboratory tests may be conducted.

# Biological evaluation of medical devices —

## Part 5: Tests for *in vitro* cytotoxicity

### 1 Scope

This part of ISO 10993 describes test methods to assess the *in vitro* cytotoxicity of medical devices.

These methods specify the incubation of cultured cells either directly or through diffusion

- a) with extracts of a device, and/or
- b) in contact with a device.

These methods are designed to determine the biological response of mammalian cells *in vitro* using appropriate biological parameters.

### 2 Normative references

The following normative documents contain provisions which, through reference in this text, constitute provisions of this part of ISO 10993. For dated references, subsequent amendments to, or revisions of, any of these publications do not apply. However, parties to agreements based on this part of ISO 10993 are encouraged to investigate the possibility of applying the most recent editions of the normative documents indicated below. For undated references, the latest edition of the normative document referred to applies. Members of ISO and IEC maintain registers of currently valid International Standards.

ISO 10993-1, *Biological evaluation of medical devices — Part 1: Evaluation and testing*.

ISO 10993-12:1996, *Biological evaluation of medical devices — Part 12: Sample preparation and reference materials*.

### 3 Terms and definitions

For the purposes of this part of ISO 10993, the terms and definitions given in ISO 10993-1 and the following apply.

#### 3.1

##### **negative control material**

material which, when tested in accordance with this part of ISO 10993, does not produce a cytotoxic response

**NOTE** The purpose of the negative control is to demonstrate background response. For example, high-density polyethylene<sup>1)</sup> for synthetic polymers, and aluminium oxide ceramic rods for dental material, have been used as negative controls.

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1) High-density polyethylene can be obtained from the U.S. Pharmacopeia (Rockville, Maryland, USA) and Food and Drug Safety Center, Hatano Research Institute (Ochiai 729-5, Hadanoshi, Kanagawa 257 - Japan). This information is given for the convenience of the user of this part of ISO 10993 and does not constitute an endorsement by ISO of these products.

### 3.2

#### **positive control material**

material which, when tested in accordance with this part of ISO 10993, provides a reproducible cytotoxic response

NOTE The purpose of the positive control is to demonstrate appropriate test system response. For example, an organo-tin stabilized poly(vinylchloride)<sup>2)</sup> has been used as a positive control for solid materials and extracts. Dilutions of phenol, for example, have been used as a positive control for extracts.

### 3.3

#### **reagent control**

extraction vehicle without test material subjected to extraction conditions and test procedures

NOTE For the purposes of this part of ISO 10993, this definition replaces that given in 3.1 of ISO 10993-12:1996.

### 3.4

#### **culture vessels**

vessels appropriate for cell culture, including glass Petri dishes, plastic culture flasks or plastic multiwells and microtitre plates

NOTE These vessels may be used interchangeably in these methods provided that they meet the requirements of tissue culture grade and are suitable for use with mammalian cells.

### 3.5

#### **subconfluency**

approximately 80 % confluency, i.e. the end of the logarithmic phase of growth

## 4 Sample preparation

### 4.1 General

The test shall be performed on either

- a) an extract of the material; and/or
- b) the material itself.

Sample preparation shall be in accordance with ISO 10993-12.

### 4.2 Preparation of liquid extracts of material

#### 4.2.1 Principles of extraction

Extraction conditions should attempt to simulate or exaggerate the conditions of clinical use so as to determine the potential toxicological hazard, without causing significant changes in the test material such as fusion, melting or alteration of the chemical structure.

NOTE The concentration of any endogenous or extraneous substances in the extract, and hence the amount of exposure to the test cells, depends on the interfacial area, extraction volume, pH, chemical solubility, diffusion rate, osmolarity, agitation, temperature, time and other factors.

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2) Organo-tin poly(vinyl chloride) positive control material is available from SIMS Portex Ltd, Hythe, Kent, CT21 6JL, UK, (product No 499-300-000). The ZDEC and ZDBC polyurethanes are available from Food and Drug Safety Center, Hatano Research Institute, (Ochiai 729-5, Hadanoshi, Kanagawa 257, Japan). This information is given for the convenience of the user of this part of ISO 10993 and does not constitute an endorsement by ISO of the product.

#### 4.2.2 Extraction vehicle

For mammalian cell assays, one or more of the following solvents shall be used. The choice of the extraction vehicle(s) shall be justified:

- a) culture medium with serum;
- b) culture medium without serum;
- c) physiological saline solution;
- d) other suitable solvent.

NOTE The choice of solvent should reflect the aim of the extraction and consideration shall be given to the use of both a polar and a nonpolar solvent. Suitable solvents include purified water, vegetable oil and dimethyl sulfoxide (DMSO). DMSO is known to be cytotoxic in selected assay systems at greater than 0,5 % (volume fraction) concentrations.

#### 4.2.3 Extraction conditions

**4.2.3.1** The extraction shall be performed in sterile, chemically inert closed containers using aseptic techniques, in general accordance with ISO 10993-12.

**4.2.3.2** Recommended extraction conditions are:

- a) not less than 24 h at  $(37 \pm 2)$  °C;
- b)  $(72 \pm 2)$  h at  $(50 \pm 2)$  °C;
- c)  $(24 \pm 2)$  h at  $(70 \pm 2)$  °C;
- d)  $(1 \pm 0,2)$  h at  $(121 \pm 2)$  °C.

The recommended conditions may be applied according to the device characteristics and specific conditions for use.

Extraction procedures using culture medium with serum can only be used under the conditions specified in 4.2.3.2 a).

**4.2.3.3** If the extract is filtered, centrifuged or processed by other methods prior to being applied to the cells, this shall be included in the final report (see clause 9). Any pH adjustment of the extract shall be reported. Manipulation of the extract, such as by pH adjustment, could influence the result.

### 4.3 Preparation of material for direct-contact tests

**4.3.1** Materials that have various shapes, sizes or physical states (i.e. liquid or solid) may be tested without modification in the cytotoxicity assays.

The preferred sample of a solid specimen should have at least one flat surface.

Adjustments shall be made for other shapes and physical states.

**4.3.2** Sterility of the test specimen shall be taken into account.

**4.3.2.1** Test materials from sterilized devices shall be handled aseptically throughout the test procedure.

**4.3.2.2** Test materials from devices which are normally supplied non-sterile but are sterilized before use shall be sterilized by the method recommended by the manufacturer and handled aseptically throughout the test procedure.

The effect of sterilization methods or agents on the device should be considered in defining the preparation of the test material prior to use in the test system.

**4.3.2.3** Test materials from devices not required to be sterile in use shall be used as supplied and handled aseptically throughout the test procedure.

**4.3.3** Liquids shall be tested by either

- a) direct deposition; or
- b) deposition onto a biologically inert absorbent matrix.

NOTE Filter discs have been found to be suitable.

**4.3.4** If appropriate, materials classed as superabsorbent shall be soaked with culture medium prior to testing to prevent sorption of the culture medium in the testing vessel.

## 5 Cell lines

**5.1** Established cell lines are preferred and where used shall be obtained from recognized repositories<sup>3)</sup>.

**5.2** Where specific sensitivity is required, primary cell cultures, cell lines and organo-typic cultures obtained directly from living tissues shall only be used if reproducibility and accuracy of the response can be demonstrated.

**5.3** If a stock culture of a cell line is stored, storage shall be at  $-80\text{ }^{\circ}\text{C}$  or below in the corresponding culture medium but containing a cryoprotectant, e.g. dimethylsulfoxide or glycerol. Long-term storage (several months up to many years) is only possible at  $-130\text{ }^{\circ}\text{C}$  or below.

**5.4** Only cells free from mycoplasma shall be used for the test. Before use, stock cultures should be tested for the absence of mycoplasma.

## 6 Culture medium

**6.1** The culture medium shall be sterile.

**6.2** The culture medium with or without serum shall meet the growth requirements of the selected cell line.

NOTE Antibiotics may be included in the media provided that they do not adversely affect the assays.

The stability of the culture medium varies with the composition and storage conditions. Media containing serum and glutamine shall be stored at  $2\text{ }^{\circ}\text{C}$  to  $8\text{ }^{\circ}\text{C}$  for no more than one week. Glutamine-containing media without serum shall be stored at  $2\text{ }^{\circ}\text{C}$  to  $8\text{ }^{\circ}\text{C}$  for no more than two weeks.

**6.3** The culture medium shall be maintained at a pH of between 7.2 and 7.4.

## 7 Preparation of cell stock culture

**7.1** Using the chosen cell line and culture medium, prepare sufficient cells to complete the test. If the cells are to be grown from cultures taken from storage, remove the cryoprotectant, if present. Subculture the cells at least once before use.

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<sup>3)</sup> For example, recommended cell lines are American Type Culture Collection CCL 1 (NCTC clone 929), CCL 163 (Balb/3T3 clone A31), CCL 171 (MRC-5) and CCL 75 (WI-38), CCL 81 (Vero) and CCL 10 [(BHK-21 (C-13)] and V-79 379A. This information is given for the convenience of the user of this part of ISO 10993 and does not constitute an endorsement by ISO of the products named. Other cell lines may be used if they can be shown to lead to the same or more relevant results.

**7.2** Remove and resuspend the cells by enzymatic and/or mechanical disaggregation, using a method appropriate for the cell line.

## 8 Test procedures

### 8.1 Number of replicates

A minimum of three replicates shall be used for test samples and controls.

### 8.2 Test on extracts

**8.2.1** This test allows both qualitative and quantitative assessment of cytotoxicity.

**8.2.2** Pipette an aliquot of the continuously stirred cell suspension into each of a sufficient number of vessels for exposure to the extracts. Distribute the cells evenly over the surface of each vessel by gentle rotation.

**8.2.3** Incubate the cultures at  $(37 \pm 2)$  °C in air with or without 5 % (volume fraction) carbon dioxide as appropriate for the buffer system chosen for the culture medium.

The test should be performed on a subconfluent monolayer or on freshly suspended cells.

In the colony-forming assay, only an appropriate low cell density shall be used.

**8.2.4** Verify the subconfluency and the morphology of the cultures with a microscope before starting the test.

**8.2.5** Perform the test on

- a) the original extract; and
- b) a dilution series of the extract, using the culture medium as diluent.

If monolayers are used for the test, remove and discard the culture medium from the cultures and add an aliquot of the extract or dilution thereof into each of the vessels.

If suspended cells are used for the test, add the extract or dilution thereof into each of the replicate vessels immediately after preparation of the cell suspension.

**8.2.6** When a non-physiological extract is used, e.g. water, the extract shall be tested at the highest physiologically compatible concentration after dilution in culture medium.

NOTE Concentrated culture medium, e.g. 2×, 5×, is recommended for use in diluting aqueous extracts.

**8.2.7** Add known aliquots of the reagent blank and the negative and positive controls into additional replicate vessels.

NOTE A fresh culture medium control may also be tested, if appropriate.

**8.2.8** Incubate the vessels, using the same conditions as described in 8.2.3, for an appropriate interval corresponding to the selected specific assay.

**8.2.9** After an incubation period of at least 24 h, determine the cytotoxic effects in accordance with 8.5.

### 8.3 Test by direct contact

**8.3.1** This test allows both qualitative and quantitative assessment of cytotoxicity.

**8.3.2** Pipette a known aliquot of the continuously stirred cell suspension into each of a sufficient number of vessels for direct exposure to the test sample. Distribute the cells evenly over the surface of each vessel by gentle horizontal rotation.

**8.3.3** Incubate the culture at  $(37 \pm 2)$  °C in air with or without 5 % (volume fraction) carbon dioxide as appropriate for the buffer system chosen for the culture medium until the cultures have grown to subconfluency.

**8.3.4** Verify the subconfluency and the morphology of the cultures with a microscope before starting the test.

**8.3.5** Remove and discard the culture medium. Then add fresh culture medium to each vessel.

**8.3.6** Carefully place individual specimens of the test sample on the cell layer in the centre of each replicate vessel. Ensure that the specimen covers approximately one-tenth of the cell layer surface.

Exercise care to prevent unnecessary movement of the specimens, as this could cause physical trauma to the cells, for example patches of dislodged cells.

NOTE When appropriate, the specimen can be placed in the culture vessel prior to the addition of the cells.

**8.3.7** Prepare replicate vessels for both the negative-control and positive-control materials.

**8.3.8** Incubate the vessels, under the same conditions as in 8.3.3, for an appropriate interval (a minimum of 24 h) corresponding to the selected specific assay.

**8.3.9** Discard the supernatant culture medium and determine the cytotoxic effects in accordance with 8.5.

## **8.4 Test by indirect contact**

### **8.4.1 Agar diffusion test**

**8.4.1.1** This test allows a qualitative assessment of cytotoxicity. This assay is not appropriate for leachables that cannot diffuse through the agar layer, or that should react with agar.

**8.4.1.2** Pipette a known aliquot of the continuously stirred cell suspension into each of a sufficient number of replicate vessels for the test. Distribute the cells evenly over the surface of each vessel by gentle horizontal rotation.

**8.4.1.3** Incubate the cultures at  $(37 \pm 2)$  °C in air with or without 5 % (volume fraction) carbon dioxide as appropriate for the buffer system chosen for the culture medium until the cultures have grown to approximate confluence at the end of the logarithmic phase of the growth curve.

**8.4.1.4** Verify the subconfluency and the morphology of the cultures with a microscope before starting the test.

**8.4.1.5** Remove and discard the culture medium from the vessel. Then mix fresh culture medium containing serum with melted agar to obtain a final mass concentration of agar of 0,5 % to 2 %, and pipette an appropriate volume into each vessel. Use only agar that is suitable for the growth of mammalian cells in culture. The agar culture medium mixture should be in a liquid state and at a temperature that is compatible with mammalian cells.

NOTE Agar is available in various molecular weight ranges and purities.

**8.4.1.6** Carefully place replicate specimens of the test sample on the solidified agar layer in each vessel. Ensure that the specimen covers approximately one-tenth of the cell layer surface.

Presoak any absorbent material with the culture medium before placing it on the agar to prevent dehydration of the agar.

**8.4.1.7** Prepare replicate vessels with both negative-control and positive-control specimens.

**8.4.1.8** Incubate the vessels, using the same conditions as described in 8.4.1.3, for between 24 h and 72 h.

**8.4.1.9** Examine the cells to determine cytotoxic effects before and after carefully removing the specimens from the agar.

Use of a vital stain, e.g. neutral red, may aid in the detection of cytotoxicity. The vital stain may be added before or after the incubation with the specimen. If the stain is added before the incubation, protect the cultures from light to prevent cell damage elicited by photoactivation of the stain.

## 8.4.2 Filter diffusion test

**8.4.2.1** This test allows a qualitative assessment of cytotoxicity.

**8.4.2.2** Place a surfactant-free filter of pore size 0,45 µm into each vessel, and add a known aliquot of the continuously stirred cell suspension into each of a sufficient number of replicate vessels for the test. Distribute the cells evenly over the surface of each filter by gentle rotation.

**8.4.2.3** Incubate the cultures at  $(37 \pm 2)$  °C in air with or without 5 % (volume fraction) carbon dioxide as appropriate for the buffer system chosen for the culture medium until the cultures have grown to approximate confluence at the end of the logarithmic phase of the growth curve.

**8.4.2.4** Verify the subconfluency and the morphology of the cultures with a microscope before starting the test.

**8.4.2.5** Remove and discard the culture medium from the vessels. Then transfer the filters, cell-side down, on to a layer of solidified agar (see 8.4.1.6).

**8.4.2.6** Carefully place the replicate specimens of the test sample on the acellular (top) side of the filter. Retain liquid extracts and freshly mixed compounds in non-reactive rings placed on the filter.

**8.4.2.7** Prepare replicate filters with both the negative-control and positive-control specimens.

**8.4.2.8** Incubate the vessels, using the same conditions described in 8.4.2.3, for  $2 \text{ h} \pm 10 \text{ min}$ .

**8.4.2.9** Carefully remove the specimens from the filter and carefully separate the filter from the agar surface.

**8.4.2.10** Determine the cytotoxic effects using an appropriate stain procedure.

## 8.5 Determination of cytotoxicity

**8.5.1** Determine cytotoxic effects by either qualitative or quantitative means.

- a) **Qualitative evaluation:** examine the cells microscopically, using cytochemical stain if desired. Assess changes in, for example, general morphology, vacuolization, detachment, cell lysis and membrane integrity. The change from normal morphology shall be recorded in the test report descriptively or numerically. A useful way to grade test materials is presented below.

Cytotoxicity scale	Interpretation
0	Noncytotoxic
1	Mildly cytotoxic
2	Moderately cytotoxic
3	Severely cytotoxic

The method of evaluation and the results of the evaluation shall be included in the test report.

- b) **Quantitative evaluation:** measure the parameter cell death, inhibition of cell growth, cell proliferation or colony formation. The number of cells, amount of protein, release of enzymes, release of vital dye, reduction of vital dye or other measurable parameter may be quantified by objective means. The objective measure and response shall be recorded in the test report.

NOTE For particular methods of determining cytotoxicity, a zero time or baseline cell culture control may be necessary.